Installation of The M3 System® in an Air Rotation Unit for Proactive Pandemic Protection

Location: 750,000 Sq Ft Textile Distribution Center
North Carolina, USA

Client: PVH Corporation

As one of the world's leading importers of RMG, *Ready Made Goods,* the company has a corporate philosophy of building on their core strengths. Their priorities are to deliver sustainable growth through aspirational products, support their pillars of corporate responsibility to advance the communities where we work and live, and develop a talented and skilled workforce that embodies our values.

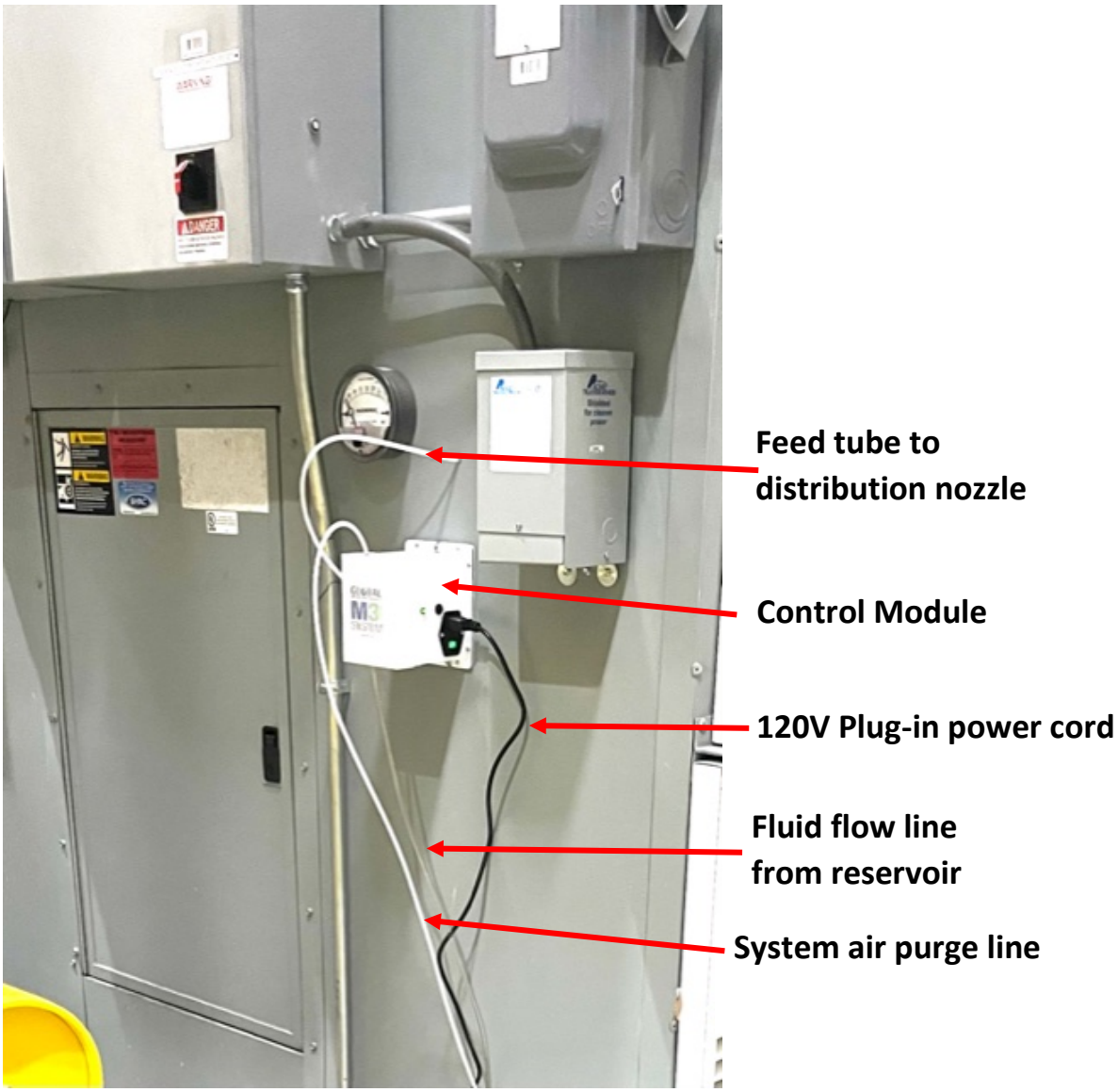
The company has over 40,000 associates operating in more than 40 countries and generated $9.9 billion in revenues in 2019. A prime ambition is for all products and business operations to generate zero waste, zero carbon emissions and zero hazardous chemicals.

As all companies today, this company recognizes that the health and wellbeing of its employees is the bedrock foundation of corporate responsibility.
They have put materials and methods in place to ensure a safe indoor environment both domestically and overseas for many years and have now embraced the most innovative, ground breaking technology available.
Some initial results of that technology are included herein for your review.

www.m3systemprotection.com

Subject: M3 System install in ARU #9.

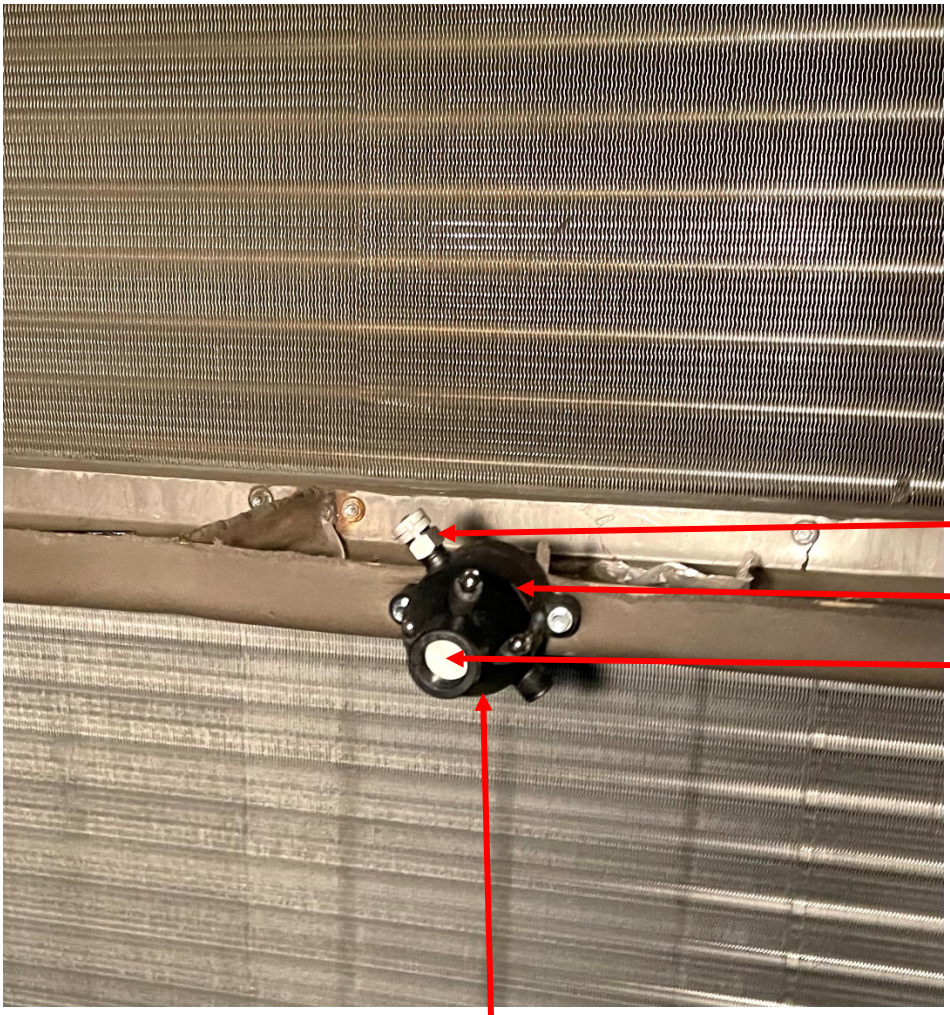
The M3 System® was installed on a single ARU, *Air Rotation Unit,* designated as ARU #9. It should be noted that there is a total of nine (9) ARUs located in the contiguous space. Unit #9 was in a somewhat remote location but none the less exposed to the general distribution open environment. This would allow air from the rest of the distribution space to potentially mix with the air in this location.



The entire system was installed in less than one (1) hour. Total system weight is less than 5 kg. Fully automatic once activated. At no time are the unit system components tapped into, breached or reconfigured.

Pre M3 System® activation on 01/14/2021 results indicated a total of 1,100 individual spores broken down into three (3) varieties. Each of the varieties identified and quantified are, in fact, allergenic and potentially toxigenic based on individual occupant medical history and individual underlying chronic conditions.

Post M3 System® installation sampling on 01/19/2021 resulted in a significant drop in total spore count with only five (5) days of operation of The M3 System®. Total spore count was quantified at 110 spores. In addition to the drop of approximately 90% in total count is should be noted that one individual spore, Cladosporium, was totally removed.



Dispersion Nozzle mounted ready for connection to Control Module

On Pre-M3 System® installation several samples were also collected at the are around ARU #7. This unit was operating without benefit of The M3 System® and indicated the presence of two (2) allergenic and potentially toxigenic spores present. The presence of pathogenic bio-aerosols here is indicative of the fact that the enclosed space is very large and these types of items can be and are entrained in other parts of the facility which would migrate to the area of ARU #7.

The results of the 01/19/2021 sampling indicates that the single unit installed in ARU #9 is making the air in that location safer for occupants and yet attacking other pathogen laden air migrating to its space location.

Current practice of decontaminating the ARUs is a fogging procedure undertaken monthly. This methodology does not track pathogen buildup or bioburden accumulation and the ability to mitigate it "as it happens." Spore counts could increase dramatically and unknowingly without daily testing. Installation of The M3 System® provides a steady, constant treatment of the air.

PVH's corporate mission of providing a safe and healthy indoor environment for staff, workers and visitors has always been a commitment and embracing new technology, once vetted, proves that PVH lives up to its commitments. We are proud the be a part of that in a small way.

CONTROL MODULE

Our proprietary sequence of operation is built into a compact pumping and control module that can be installed inside your air handler, on the exterior or on an adjacent wall. This module is designed to input only authorized organic based products effective against harmful pathogens. Our proprietary sequencing of frequency and amount of product input has been scientifically tested in more than 4,000 buildings worldwide. We assure you of maximum results with minimum costs.

DISPERSION NOZZLE

Based on the micron size output we require to maximize the Brownian Theory of Motion effect, our dispersion nozzle was designed and selected to work only with our component package. There are multiple options for mounting the dispersion nozzle based on your particular air handler configuration.

FOOT VALVE

To ensure a proper fluid pickup and delivery to the pumping and control module our engineers designed the Foot Valve with a built-in weight to keep it level for maximum fluid pickup. It has been designed with no moving parts for trouble free operation.

THE SCIENCE

Utilizing the Brownian Theory of Motion, defined as, “The erratic random movement of microscopic particles in a fluid, as a result of continuous bombardment from molecules of the surrounding medium” our team has developed a way to use this principal to micro-infuse millions of molecules of our organic,

extremely high efficacy, botanical based product into and onto the surfaces of the buildings HVAC system so that any pathogenic bioaerosols such as viruses, fungi, bacteria, yeasts and non-viable particulates such as pollen will come into contact with each other through recirculation of the air in the occupied space through the confines of the HVAC system.

When a molecule of high efficacy contacts a unit of viability such as a virus, fungal, bacteria or yeast the viable unit's ability to reproduce is disrupted and destroyed. Similarly, non-viable items such as pollen have their capability to be allergenic negated.

The average adult breathes in excess of 50 cubic meters of air per 24-hour period. Dry air is composed of approximately 78% nitrogen, 21% oxygen, 1% argon, carbon dioxide, water vapor and a minute number of other gases. Non controllable factors contribute to that healthy air also containing a very large variable amounts of viruses, fungi, bacteria, yeasts, plant pollen and numerous other potentially harmful disease carrying particulates.

The key to keeping building occupants safe and healthy is the ability to provide Proactive Pandemic Protection automatically, constantly and safely with innovative technology that is "Organic based, Non-GMO, Tested, Proven and Approved."

Products used proven effective against COVID-19 SARS (CoV2) by USA laboratory approved by CDC

Arthur V. Martin Ph.D. President
Principal Research Scientist

Arthur V. Martin Ph.D.

Tel: 1.843-368-7063

amartin@gicclc.com

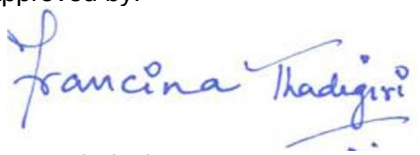
eurofins | EMLab P&K

Report for:

Mr. Lou Schwartz
PVH Corp
1000 Quality Way
Jonesville, NC 28642

Regarding: Project: JONESVILLE ARU'S TESTING; CASSETTES
EML ID: 2557764

Approved by:



Technical Manager
Francina Thadigiri

Dates of Analysis:
Spore trap analysis: 01-15-2021

Service SOPs: Spore trap analysis (EM-MY-S-1038)
AIHA-LAP, LLC accredited service, Lab ID #179623

All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank correction of results is not applied. The results relate only to the samples as received. Sample air volume is supplied by the client.

Eurofins EMLab P&K ("the Company") shall have no liability to the client or the client's customer with respect to decisions or recommendations made, actions taken or courses of conduct implemented by either the client or the client's customer as a result of or based upon the Test Results. In no event shall the Company be liable to the client with respect to the Test Results except for the Company's own willful misconduct or gross negligence nor shall the Company be liable for incidental or consequential damages or lost profits or revenues to the fullest extent such liability may be disclaimed by law, even if the Company has been advised of the possibility of such damages, lost profits or lost revenues. In no event shall the Company's liability with respect to the Test Results exceed the amount paid to the Company by the client therefor.

Eurofins EMLab P&K's LabServe® reporting system includes automated fail-safes to ensure that all AIHA-LAP, LLC quality requirements are met and notifications are added to reports when any quality steps remain pending.

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

Client: PVH Corp
C/O: Mr. Lou Schwartz
Re: JONESVILLE ARU'S TESTING; CASSETTES

Date of Sampling: 01-14-2021
Date of Receipt: 01-15-2021
Date of Report: 01-18-2021

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	1: ARU #9		2: ARU #7		3: OUTSIDE	
Comments (see below)	A		None		None	
Lab ID-Version‡:	12194000-1		12194002-1		12194004-1	
Analysis Date:	01/15/2021		01/15/2021		01/15/2021	
	raw ct.	spores/m3	raw ct.	spores/m3	raw ct.	spores/m3
Ascospores						
Basidiospores	4	210			63	3,400
Bipolaris/Drechslera group						
Botrytis						
Chaetomium						
Cladosporium	1	53	1	53	1	53
Curvularia						
Epicoccum						
Fusarium						
Myrothecium						
Nigrospora						
Other colorless						
Penicillium/Aspergillus types†	23	790	5	270	2	110
Pithomyces						
Rusts						
Smuts, Periconia, Myxomycetes						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Zygomycetes						
Background debris (1-4+)††	2+		2+		2+	
Hyphal fragments/m3	13		13		< 13	
Pollen/m3	< 13		< 13		< 13	
Skin cells (1-4+)	< 1+		< 1+		< 1+	
Sample volume (liters)	75		75		75	
§ TOTAL SPORES/m3		1,100		320		3,500

Comments: A) 11 of the raw count *Penicillium/Aspergillus* type spores were present as a single clump.

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.
† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium, Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.
††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m^3 divided by the raw count, expressed in spores/m^3. The limit of detection is the analytical sensitivity (in spores/m^3) multiplied by the sample volume (in liters) divided by 1000 liters.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.
‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".
§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.

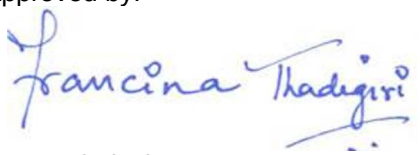
eurofins | EMLab P&K

Report for:

Mr. Lou Schwartz
PVH Corp
1000 Quality Way
Jonesville, NC 28642

Regarding: Project: JONESVILLE ARU'S TESTING; CASSETTES
EML ID: 2557764

Approved by:



Technical Manager
Francina Thadigiri

Dates of Analysis:
Spore trap analysis other particles-Supplement: 01-15-2021

Service SOPs: Spore trap analysis other particles-Supplement (EM-MY-S-1038)
AIHA-LAP, LLC accredited service, Lab ID #179623

All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank correction of results is not applied. The results relate only to the samples as received. Sample air volume is supplied by the client.

Eurofins EMLab P&K ("the Company") shall have no liability to the client or the client's customer with respect to decisions or recommendations made, actions taken or courses of conduct implemented by either the client or the client's customer as a result of or based upon the Test Results. In no event shall the Company be liable to the client with respect to the Test Results except for the Company's own willful misconduct or gross negligence nor shall the Company be liable for incidental or consequential damages or lost profits or revenues to the fullest extent such liability may be disclaimed by law, even if the Company has been advised of the possibility of such damages, lost profits or lost revenues. In no event shall the Company's liability with respect to the Test Results exceed the amount paid to the Company by the client therefor.

Eurofins EMLab P&K's LabServe® reporting system includes automated fail-safes to ensure that all AIHA-LAP, LLC quality requirements are met and notifications are added to reports when any quality steps remain pending.

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

Client: PVH Corp
C/O: Mr. Lou Schwartz
Re: JONESVILLE ARU'S TESTING; CASSETTES

Date of Sampling: 01-14-2021
Date of Receipt: 01-15-2021
Date of Report: 01-18-2021

OTHER BIOLOGICAL PARTICLES REPORT: NON-VIABLE METHODOLOGY

Location:	1: ARU #9		2: ARU #7		3: OUTSIDE	
Comments (see below)	None		None		None	
Lab ID-Version‡:	12194001-1		12194003-1		12194005-1	
	raw ct.	particles/m3	raw ct.	particles/m3	raw ct.	particles/m3
POLLEN						
Eucalyptus (Eucalyptus)						
Grass (Poaceae)						
Mulberry (Morus)						
Oak (Quercus)						
Other						
Pine (Pinaceae)						
Ragweed (Ambrosieae)						
Sycamore (Platanus)						
OTHER PLANT						
Algae						
Diatoms						
Fern, moss, etc. spores						
Other (wood, trichomes, etc.)						
OTHER PARTICLES:						
ANIMAL						
Epithelial (skin) cells	9	120	18	240	7	93
Hair						
Insect parts						
Mites						
FUNGI						
Hyphal fragments	1	13	1	13		
NON-BIOLOGICAL						
Cellulose fibers	6	80	7	93	1	13
Glass fiber						
Starch particles	1	13	1	13		
Synthetic fibers			4	53	1	13
Background debris (1-4+)†	2+		2+		2+	
Sample volume (liters)	75		75		75	

Comments:

The analytical sensitivity is the spores/m3 divided by the raw count. The limit of detection is the analytical sensitivity multiplied by the sample volume divided by 1000.

Carbonaceous particles include soot and other combustion products. In most instances a detailed analysis of soot can be accomplished using scanning electron microscopy.

Note: Interpretation is left to the company and/or persons who conducted the field work.

† Background debris is an indication of the amounts of non-biological particulate matter present on the slide (dust in the air) and is graded from 1+ to 4+ with 4+ indicating the largest amounts. To evaluate dust levels it is important to account for differences in sample volume.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

Client: PVH Corp
C/O: Mr. Lou Schwartz
Re: JONESVILLE ARU'S TESTING; CASSETTES

Date of Sampling: 01-14-2021
Date of Receipt: 01-15-2021
Date of Report: 01-18-2021

MoldRANGE™, Local Climate; Extended Outdoor Comparison

Outdoor Location: 3, OUTSIDE

Fungi Identified	Outdoor data	Typical Outdoor Data for: January in Southeast† EMLab Regional Climate code¹ B Annual Temp, A Elev., B Rain, A Temp. Range (n‡=342)						Typical Outdoor Data for: The entire year in Southeast† EMLab Regional Climate code¹ B Annual Temp, A Elev., B Rain, A Temp. Range (n‡=5402)					
Project zip code 28642	spores/m3	very low	low	med	high	very high	freq %	very low	low	med	high	very high	freq %
Generally able to grow indoors*													
Alternaria	-	7	7	13	20	42	13	13	13	27	67	110	43
Bipolaris/Drechslera group	-	-	-	-	-	-	4	7	13	13	40	67	21
Chaetomium	-	-	-	-	-	-	4	7	7	13	33	67	5
Cladosporium	53	33	53	190	590	1,100	70	80	160	690	2,100	3,600	91
Curvularia	-	7	7	13	13	26	6	7	13	27	53	110	30
Nigrospora	-	-	-	-	-	-	4	7	8	13	27	53	20
Penicillium/Aspergillus types	110	40	53	160	380	590	72	53	100	270	750	1,300	79
Stachybotrys	-	-	-	-	-	-	< 1	7	13	13	43	140	2
Torula	-	-	-	-	-	-	2	7	13	13	39	53	11
Seldom found growing indoors**													
Ascospores	-	27	53	110	430	1,400	70	53	110	590	2,300	4,200	91
Basidiospores	3,400	53	130	710	4,000	8,500	95	210	590	3,500	15,000	30,000	98
Rusts	-	-	-	-	-	-	4	7	13	20	53	110	23
Smuts, Periconia, Myxomycetes	-	13	13	20	42	86	43	13	19	53	130	230	72
§ TOTAL SPORES/m3	3,500												

¹EMLab Regional Climate codes are a climate classification scheme for regional geographic areas containing multiple states. The MoldRANGE™ Local Climate report uses the sampling location zip code to identify the EMLab Regional Climate code in that area. Using information available from the NOAA weather database, the EMLab Regional Climate code sharpens the precision of the MoldRANGE™reporting system, providing more reliable estimates of the range and average concentrations of the different airborne fungal spore types for each region. Additional information on the EMLab Regional Climate code system can be found on the last page of this report.

†The Typical Outdoor Data represents the typical outdoor spore levels across the region's group of states for the time period and EMLab Regional Climate code indicated. The last column represents the frequency of occurrence. The very low, low, med, high, and very high values represent the 10, 20, 50, 80, and 90 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 20% of the time it is present in levels above the detection limit and below 53 spores/m3. These values are updated periodically and if not enough data is available to make a statistically meaningful assessment, it is indicated with a dash.

‡ n is the sample size used to calculate the MoldRANGE™ Local Climate data summarized in the table.

* The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

** These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

Client: PVH Corp
C/O: Mr. Lou Schwartz
Re: JONESVILLE ARU'S TESTING; CASSETTES

Date of Sampling: 01-14-2021
Date of Receipt: 01-15-2021
Date of Report: 01-18-2021

Understanding EMLab Regional Climate Codes

Outdoor airborne spore concentrations are strongly influenced by climate and weather patterns, often resulting in pronounced seasonal and diurnal cycles (Burge 1995). The seasonal climatic changes directly affect the growth cycle of plants, thereby influencing fungal growth, spore maturation, and release cycles. By evaluating outdoor spore concentrations across similar climatic zones rather than for the state as a whole, it is possible to provide a more representative estimate of typical outdoor spore levels and frequency of occurrence for different airborne fungal spore types in a given area.

The EMLab Regional Climate code system is a novel classification system that uses data from the NOAA - National Oceanic and Atmospheric Administration database to define unique climate zones. The following climate variables, for each regional zip code, are obtained from NOAA and assigned a letter code of A (above the regional average for that variable) or B (below the regional average for that variable):

1. Annual High Temperature
2. Elevation
3. Rainfall/Precipitation
4. Monthly Temperature Range

The result is a 4-character code assigned to each statewide zip code, referred to as the Regional Climate Code. Below are some examples of decoded Regional Climate Codes:

AAAA = Above avg. Annual High Temperature, Above avg. Elevation, Above avg. Rainfall/Precipitation, Above avg. Monthly Temperature Range
AABB = Above avg. Annual High Temperature, Above avg. Elevation, Below avg. Rainfall/Precipitation, Below avg. Monthly Temperature Range
BBAA = Below avg. Annual High Temperature, Below avg. Elevation, Above avg. Rainfall/Precipitation, Above avg. Monthly Temperature Range

The actual outdoor air sample data from matching regional climate codes in each group of states are then compiled in a manner relating typical spore concentrations and frequency of occurrence.

The data presented in this report is from the Southeast Region which includes the states of: AL, FL, GA, NC, SC, and VA

The NOAA regional climate variables were selected by mapping data points from a subset of approximately 145,000 weather and geographic database entries to over 80,000 outdoor spore trap samples with known zip codes and assessing them using orthogonal array experimental design techniques. The results were then compared to the typical ranges of spore types found when grouping zip codes using the Koppen-Geiger climatic classification system; a commonly used climatic system that provides an objective numerical definition in terms of climatic elements such as temperature, rainfall, and other seasonal characteristics . The EMLab Regional Climate codes showed improved granularity and refinement of the zip code groupings, implying a better representation of the expected range of spore types to be found within an individual zip code.

The values on this report were calculated by obtaining the four variables listed above from the over 585 million data points of weather and geographic information available in the NOAA database, and determining the frequencies and percentile values of spore types by utilizing over 180,000 Eurofins EMLab P&K outdoor spore trap samples with known zip codes.

This report groups regional zip codes in relation to these EMLab Regional Climate codes and summarizes MoldRANGE™ data by month and year within each EMLab Regional Climate code.

References:

Burge, Harriet, A. Bioaerosols: Boca Raton: Lewis Publishers, pp. 163-171, 1995.

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

Client: PVH Corp
C/O: Mr. Lou Schwartz
Re: JONESVILLE ARU'S TESTING; CASSETTES

Date of Sampling: 01-14-2021
Date of Receipt: 01-15-2021
Date of Report: 01-18-2021

MoldSTAT™: Supplementary Statistical Spore Trap Report

Outdoor Summary: 3: OUTSIDE

Species detected	Outdoor sample spores/m3 (<100 / 1K / 10K / >100K)	Typical outdoor ranges (North America)	Freq. %
Ascospores	< 13	13 - 270 - 6,400	77
Basidiospores	3,400	20 - 480 - 24,000	90
Cladosporium	53	27 - 480 - 8,300	88
Penicillium/Aspergillus types	110	13 - 210 - 2,800	64
Smuts, Periconia, Myxomycetes	< 13	7 - 53 - 1,100	67
Total	3,500		

The "Typical outdoor ranges" and "Freq. %" columns show the typical low, medium, and high spore counts per cubic meter and the frequency of occurrence for the given spore type. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values when the spore type is detected. For example, if the low value is 53 and the frequency of occurence is 63%, it would mean that we typically detect the given spore type on 63 percent of all outdoor samples and, when detected, 2.5% of the time it is present in levels below 53 spores/m3.

Indoor Samples

Location: 1: ARU #9

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: 29%	dF: 1 Result: 1.3333 Critical value: 3.8415 Inside Similar: Yes	Result: 1.0000	dF: 3 Result: 0.5000 Critical value: N/A Outside Similar: N/A	Score: 210 Result: Medium

Species Detected	Spores/m3 (<100 / 1K / 10K / >100K)
Basidiospores	210
Cladosporium	53
Penicillium/Aspergillus types	790
Total	1,100

Location: 2: ARU #7

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: 9%	dF: 1 Result: 1.3333 Critical value: 3.8415 Inside Similar: Yes	Result: 0.8000	dF: 3 Result: -0.5000 Critical value: N/A Outside Similar: N/A	Score: 141 Result: Low

Species Detected	Spores/m3 (<100 / 1K / 10K / >100K)
Cladosporium	53
Penicillium/Aspergillus types	270
Total	320

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

Client: PVH Corp
C/O: Mr. Lou Schwartz
Re: JONESVILLE ARU'S TESTING; CASSETTES

Date of Sampling: 01-14-2021
Date of Receipt: 01-15-2021
Date of Report: 01-18-2021

MoldSTAT™: Supplementary Statistical Spore Trap Report

* The Friedman chi-square statistic is a non-parametric test that examines variation in a set of data (in this case, all indoor spore counts). The null hypothesis (H0) being tested is that there is no meaningful difference in the data for all indoor locations. The alternative hypothesis (used if the test disproves the null hypothesis) is that there is a difference between the indoor locations. The null hypothesis is rejected when the result of the test is greater than the critical value. The critical value that is displayed is based on the degrees of freedom (dF) of the test and a significance level of 0.05.

** An agreement ratio is a simple method for assessing the similarity of two samples (in this case the indoor sample and the outdoor summary) based on the spore types present. A score of one indicates that the types detected in one location are the same as that in the other. A score of zero indicates that none of the types detected indoors are present outdoors. Typically, an agreement of 0.8 or higher is considered high.

*** The Spearman rank correlation is a non-parametric test that examines correlation between two sets of data (in this case the indoor location and the outdoor summary). The null hypothesis (H0) being tested is that the indoor and outdoor samples are unrelated. The alternative hypothesis (used if the test disproves the null hypothesis) is that the samples are similar. The null hypothesis is rejected when the result of the test is greater than the critical value. The critical value that is displayed is based on the degrees of freedom (dF) of the test and a significance level of 0.05.

**** MoldSCORE™ is a specialized method for examining air sampling data. It is a score between 100 and 300, with 100 indicating a greater likelihood that the airborne indoor spores originated from the outside, and 300 indicating a greater likelihood that they originated from an inside source. The Result displayed is based on the numeric score given and will be either Low, Medium, or High, indicating a low, medium, or high likelihood that the spores detected originated from an indoor source. Eurofins EMLab P&Kreserves the right to, and may at anytime, modify or change the MoldScore algorithm without notice.

Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor ranges" are based on the results of the analysis of samples delivered to and analyzed by Eurofins EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. With the statistical analysis provided, as with all statistical comparisons and analyses, false-positive and false-negative results can and do occur. Eurofins EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the data contained in, or any actions taken or omitted in reliance upon, this report.

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

Client: PVH Corp
C/O: Mr. Lou Schwartz
Re: JONESVILLE ARU'S TESTING; CASSETTES

Date of Sampling: 01-14-2021
Date of Receipt: 01-15-2021
Date of Report: 01-18-2021

MoldSCORE™: Spore Trap Report

Outdoor Sample: 3 OUTSIDE

Fungi Identified	Outdoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3
Generally able to grow indoors*			
Alternaria		ND	< 13
Bipolaris/Drechslera group		ND	< 13
Chaetomium		ND	< 13
Cladosporium		1	53
Curvularia		ND	< 13
Nigrospora		ND	< 13
Penicillium/Aspergillus types†		2	110
Stachybotrys		ND	< 13
Torula		ND	< 13
Seldom found growing indoors**			
Ascospores		ND	< 13
Basidiospores		63	3,400
Rusts		ND	< 13
Smuts, Periconia, Myxomycetes		ND	< 13
Total			**3,520**

Location: 1 ARU #9

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		1	53		102
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Penicillium/Aspergillus types†		23	790		210
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		ND	< 13		100
Basidiospores		4	210		100
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**1,053**	**Final MoldSCORE**	**210**

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

Client: PVH Corp
C/O: Mr. Lou Schwartz
Re: JONESVILLE ARU'S TESTING; CASSETTES

Date of Sampling: 01-14-2021
Date of Receipt: 01-15-2021
Date of Report: 01-18-2021

MoldSCORE™: Spore Trap Report

Location: 2 ARU #7

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		1	53		103
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Penicillium/Aspergillus types†		5	270		141
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		ND	< 13		100
Basidiospores		ND	< 13		100
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**320**	**Final MoldSCORE**	**141**

* The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

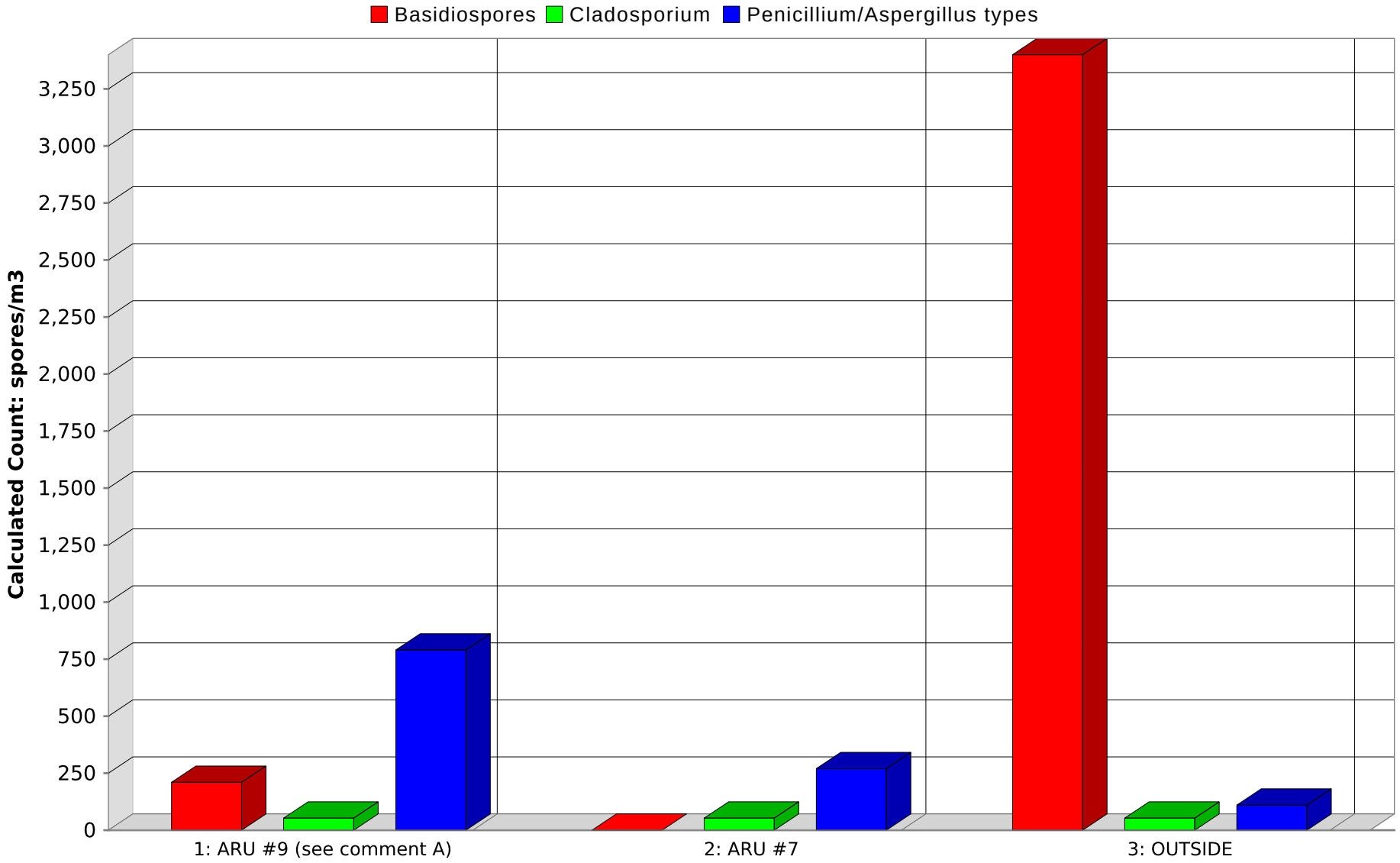
** These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

†The spores of Aspergillus and Penicillium (and others such as Acremonium, Paecilomyces) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods.

‡Rated on a scale from 100 to 300. A rating less than 150 is low and indicates a low probability of spores originating inside. A rating greater than 250 is high and indicates a high probability that the spores originated from inside, presumably from indoor mold growth. A rating between 150 and 250 indicates a moderate likelihood of indoor fungal growth. MoldSCORE is NOT intended for wall cavity samples. It is intended for ambient air samples in residences. Using the analysis on other samples (like wall cavity samples) will lead to misleading results.

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

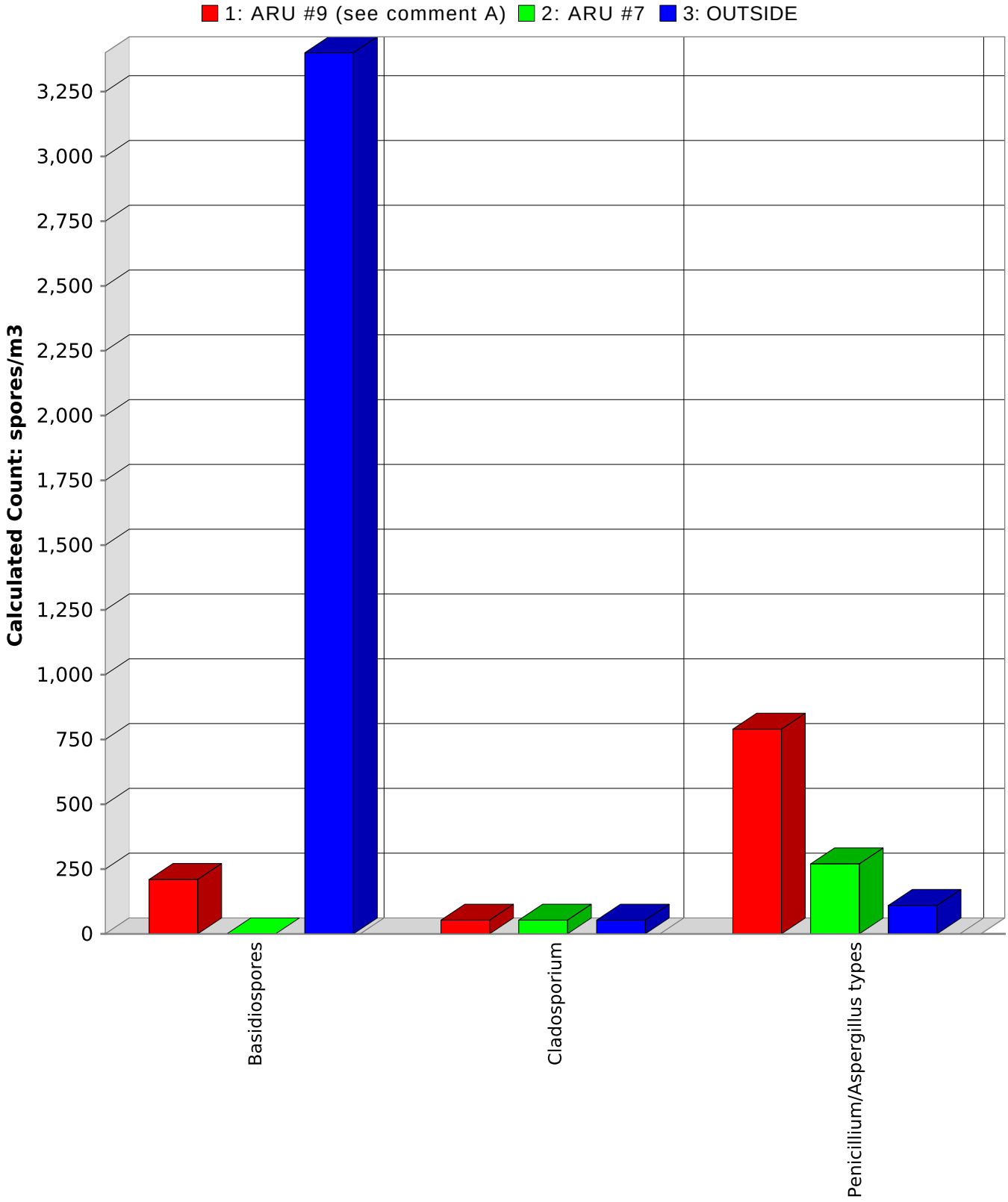


Comments: A) 11 of the raw count *Penicillium/Aspergillus* type spores were present as a single clump.

Note: Graphical output may understate the importance of certain "marker" genera.

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY



Comments: A) 11 of the raw count *Penicillium/Aspergillus* type spores were present as a single clump.

Note: Graphical output may understate the importance of certain "marker" genera.

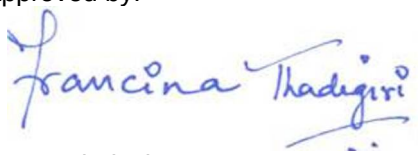
eurofins | EMLab P&K

Report for:

Mr. Lou Schwartz
PVH Corp
1000 Quality Way
Jonesville, NC 28642

Regarding: Project: JONESVILE ARU TESTING; CASSETTES
EML ID: 2560039

Approved by:



Technical Manager
Francina Thadigiri

Dates of Analysis:
Spore trap analysis: 01-20-2021

Service SOPs: Spore trap analysis (EM-MY-S-1038)
AIHA-LAP, LLC accredited service, Lab ID #179623

All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank correction of results is not applied. The results relate only to the samples as received. Sample air volume is supplied by the client.

Eurofins EMLab P&K ("the Company") shall have no liability to the client or the client's customer with respect to decisions or recommendations made, actions taken or courses of conduct implemented by either the client or the client's customer as a result of or based upon the Test Results. In no event shall the Company be liable to the client with respect to the Test Results except for the Company's own willful misconduct or gross negligence nor shall the Company be liable for incidental or consequential damages or lost profits or revenues to the fullest extent such liability may be disclaimed by law, even if the Company has been advised of the possibility of such damages, lost profits or lost revenues. In no event shall the Company's liability with respect to the Test Results exceed the amount paid to the Company by the client therefor.

Eurofins EMLab P&K's LabServe® reporting system includes automated fail-safes to ensure that all AIHA-LAP, LLC quality requirements are met and notifications are added to reports when any quality steps remain pending.

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

Client: PVH Corp
C/O: Mr. Lou Schwartz
Re: JONESVILE ARU TESTING; CASSETTES

Date of Sampling: 01-19-2021
Date of Receipt: 01-20-2021
Date of Report: 01-21-2021

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:	1: ARU #9		2: OUTSIDE	
Comments (see below)	None		None	
Lab ID-Version‡:	12206169-1		12206171-1	
Analysis Date:	01/20/2021		01/20/2021	
	raw ct.	spores/m3	raw ct.	spores/m3
Ascospores				
Basidiospores	1	53	9	480
Bipolaris/Drechslera group				
Botrytis				
Chaetomium				
Cladosporium				
Curvularia				
Epicoccum				
Fusarium				
Myrothecium				
Nigrospora				
Other colorless				
Penicillium/Aspergillus types†	1	53	1	53
Pithomyces				
Rusts				
Smuts, Periconia, Myxomycetes			1	13
Stachybotrys				
Stemphylium				
Torula				
Ulocladium				
Zygomycetes				
Background debris (1-4+)††	2+		2+	
Hyphal fragments/m3	13		40	
Pollen/m3	< 13		< 13	
Skin cells (1-4+)	< 1+		< 1+	
Sample volume (liters)	75		75	
§ TOTAL SPORES/m3		110		550

Comments:

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.
† The spores of *Aspergillus* and *Penicillium* (and others such as *Acremonium, Paecilomyces*) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.
††Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

The analytical sensitivity is the spores/m^3 divided by the raw count, expressed in spores/m^3. The limit of detection is the analytical sensitivity (in spores/m^3) multiplied by the sample volume (in liters) divided by 1000 liters.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory.
‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".
§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.

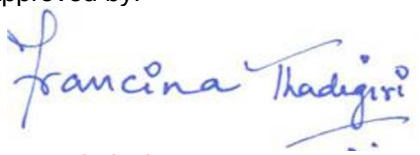
eurofins | EMLab P&K

Report for:

Mr. Lou Schwartz
PVH Corp
1000 Quality Way
Jonesville, NC 28642

Regarding: Project: JONESVILE ARU TESTING; CASSETTES
EML ID: 2560039

Approved by:



Technical Manager
Francina Thadigiri

Dates of Analysis:
Spore trap analysis other particles-Supplement: 01-20-2021

Service SOPs: Spore trap analysis other particles-Supplement (EM-MY-S-1038)
AIHA-LAP, LLC accredited service, Lab ID #179623

All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank correction of results is not applied. The results relate only to the samples as received. Sample air volume is supplied by the client.

Eurofins EMLab P&K ("the Company") shall have no liability to the client or the client's customer with respect to decisions or recommendations made, actions taken or courses of conduct implemented by either the client or the client's customer as a result of or based upon the Test Results. In no event shall the Company be liable to the client with respect to the Test Results except for the Company's own willful misconduct or gross negligence nor shall the Company be liable for incidental or consequential damages or lost profits or revenues to the fullest extent such liability may be disclaimed by law, even if the Company has been advised of the possibility of such damages, lost profits or lost revenues. In no event shall the Company's liability with respect to the Test Results exceed the amount paid to the Company by the client therefor.

Eurofins EMLab P&K's LabServe® reporting system includes automated fail-safes to ensure that all AIHA-LAP, LLC quality requirements are met and notifications are added to reports when any quality steps remain pending.

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

Client: PVH Corp
C/O: Mr. Lou Schwartz
Re: JONESVILE ARU TESTING; CASSETTES

Date of Sampling: 01-19-2021
Date of Receipt: 01-20-2021
Date of Report: 01-21-2021

OTHER BIOLOGICAL PARTICLES REPORT: NON-VIABLE METHODOLOGY

Location:	1: ARU #9		2: OUTSIDE	
Comments (see below)	None		None	
Lab ID-Version‡:	12206170-1		12206172-1	
	raw ct.	particles/m3	raw ct.	particles/m3
POLLEN				
Elm (Ulmus)				
Eucalyptus (Eucalyptus)				
Grass (Poaceae)				
Mulberry (Morus)				
Oak (Quercus)				
Other				
Pine (Pinaceae)				
Ragweed (Ambrosieae)				
Sycamore (Platanus)				
OTHER PLANT				
Algae				
Diatoms				
Fern, moss, etc. spores				
Other (wood, trichomes, etc.)			1	13
OTHER PARTICLES:				
ANIMAL				
Epithelial (skin) cells	21	280	13	170
Hair				
Insect parts			1	13
Mites				
FUNGI				
Hyphal fragments	1	13	3	40
NON-BIOLOGICAL				
Cellulose fibers	5	67	6	80
Glass fiber				
Starch particles	3	40	9	120
Background debris (1-4+)†	2+		2+	
Sample volume (liters)	75		75	

Comments:

The analytical sensitivity is the spores/m3 divided by the raw count. The limit of detection is the analytical sensitivity multiplied by the sample volume divided by 1000.

Carbonaceous particles include soot and other combustion products. In most instances a detailed analysis of soot can be accomplished using scanning electron microscopy.

Note: Interpretation is left to the company and/or persons who conducted the field work.

† Background debris is an indication of the amounts of non-biological particulate matter present on the slide (dust in the air) and is graded from 1+ to 4+ with 4+ indicating the largest amounts. To evaluate dust levels it is important to account for differences in sample volume.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

Client: PVH Corp
C/O: Mr. Lou Schwartz
Re: JONESVILE ARU TESTING; CASSETTES

Date of Sampling: 01-19-2021
Date of Receipt: 01-20-2021
Date of Report: 01-21-2021

MoldRANGE™: Extended Outdoor Comparison

Outdoor Location: 2, OUTSIDE

Fungi Identified	Outdoor data	Typical Outdoor Data for: January in North Carolina† (n‡=333)						Typical Outdoor Data for: The entire year in North Carolina† (n‡=5044)					
	spores/m3	very low	low	med	high	very high	freq %	very low	low	med	high	very high	freq %
Generally able to grow indoors*													
Alternaria	-	7	13	13	27	53	15	7	13	27	67	120	42
Bipolaris/Drechslera group	-	7	13	13	53	67	8	7	7	13	33	53	20
Chaetomium	-	-	-	-	-	-	5	7	7	13	27	70	5
Cladosporium	-	27	53	160	540	1,200	80	53	120	500	1,700	2,800	92
Curvularia	-	7	7	13	39	53	14	7	13	27	67	130	30
Nigrospora	-	7	7	13	22	53	9	7	7	13	40	53	19
Penicillium/Aspergillus types	53	27	53	130	270	530	74	53	67	200	590	1,100	77
Stachybotrys	-	-	-	-	-	-	2	7	7	13	67	120	1
Torula	-	7	7	13	32	53	7	7	7	13	47	79	10
Seldom found growing indoors**													
Ascospores	-	27	53	110	330	780	68	53	110	480	1,800	3,200	88
Basidiospores	480	53	110	480	2,700	7,200	94	130	350	2,300	10,000	20,000	98
Rusts	-	-	-	-	-	-	4	7	10	13	53	110	17
Smuts, Periconia, Myxomycetes	13	7	13	27	53	73	58	13	13	48	130	210	70
§ TOTAL SPORES/m3	550												

†The 'Typical Outdoor Data' represents the typical outdoor spore levels for the location and time frame indicated. The last column represents the frequency of occurrence. The very low, low, med, high, and very high values represent the 10, 20, 50, 80, and 90 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 20% of the time it is present in levels above the detection limit and below 53 spores/m3. These values are updated periodically, and if enough data is not available to make a statistically meaningful assessment, it is indicated with a dash.

§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.

* The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

** These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

‡n = number of samples used to calculate data.

Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor data" are based on the results of the analysis of samples delivered to and analyzed by Eurofins EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. In addition, Eurofins EMLab P&K may not have received and tested a representative number of samples for every region or time period. Eurofins EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the use or interpretation of the data contained in, or any actions taken or omitted in reliance upon, this report.

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

Client: PVH Corp
C/O: Mr. Lou Schwartz
Re: JONESVILE ARU TESTING; CASSETTES

Date of Sampling: 01-19-2021
Date of Receipt: 01-20-2021
Date of Report: 01-21-2021

MoldSTAT™: Supplementary Statistical Spore Trap Report

Outdoor Summary: 2: OUTSIDE

Species detected	Outdoor sample spores/m3 <100 1K 10K >100K	Typical outdoor ranges (North America)	Freq. %
Ascospores	< 13	13 - 270 - 6,400	77
Basidiospores	480	20 - 480 - 24,000	90
Cladosporium	< 13	27 - 480 - 8,300	88
Penicillium/Aspergillus types	53	13 - 210 - 2,800	64
Smuts, Periconia, Myxomycetes	13	7 - 53 - 1,100	67
Total	550		

The "Typical outdoor ranges" and "Freq. %" columns show the typical low, medium, and high spore counts per cubic meter and the frequency of occurrence for the given spore type. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values when the spore type is detected. For example, if the low value is 53 and the frequency of occurence is 63%, it would mean that we typically detect the given spore type on 63 percent of all outdoor samples and, when detected, 2.5% of the time it is present in levels below 53 spores/m3.

Indoor Samples

Location: 1: ARU #9

% of outdoor total spores/m3	Friedman chi-square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)
Result: 19%	dF: N/A Result: N/A Critical value: N/A Inside Similar: N/A	Result: 0.8000	dF: 3 Result: 0.8750 Critical value: N/A Outside Similar: N/A	Score: 108 Result: Low

Species Detected	Spores/m3 <100 1K 10K >100K
Basidiospores	53
Penicillium/Aspergillus types	53
Total	110

* The Friedman chi-square statistic is a non-parametric test that examines variation in a set of data (in this case, all indoor spore counts). The null hypothesis (H0) being tested is that there is no meaningful difference in the data for all indoor locations. The alternative hypothesis (used if the test disproves the null hypothesis) is that there is a difference between the indoor locations. The null hypothesis is rejected when the result of the test is greater than the critical value. The critical value that is displayed is based on the degrees of freedom (dF) of the test and a significance level of 0.05.

** An agreement ratio is a simple method for assessing the similarity of two samples (in this case the indoor sample and the outdoor summary) based on the spore types present. A score of one indicates that the types detected in one location are the same as that in the other. A score of zero indicates that none of the types detected indoors are present outdoors. Typically, an agreement of 0.8 or higher is considered high.

*** The Spearman rank correlation is a non-parametric test that examines correlation between two sets of data (in this case the indoor location and the outdoor summary). The null hypothesis (H0) being tested is that the indoor and outdoor samples are unrelated. The alternative hypothesis (used if the test disproves the null hypothesis) is that the samples are similar. The null hypothesis is rejected when the result of the test is greater than the critical value. The critical value that is displayed is based on the degrees of freedom (dF) of the test and a significance level of 0.05.

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

Client: PVH Corp
C/O: Mr. Lou Schwartz
Re: JONESVILE ARU TESTING; CASSETTES

Date of Sampling: 01-19-2021
Date of Receipt: 01-20-2021
Date of Report: 01-21-2021

MoldSTAT™: Supplementary Statistical Spore Trap Report

**** MoldSCORE™ is a specialized method for examining air sampling data. It is a score between 100 and 300, with 100 indicating a greater likelihood that the airborne indoor spores originated from the outside, and 300 indicating a greater likelihood that they originated from an inside source. The Result displayed is based on the numeric score given and will be either Low, Medium, or High, indicating a low, medium, or high likelihood that the spores detected originated from an indoor source. Eurofins EMLab P&Kreserves the right to, and may at anytime, modify or change the MoldScore algorithm without notice.

Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor ranges" are based on the results of the analysis of samples delivered to and analyzed by Eurofins EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. With the statistical analysis provided, as with all statistical comparisons and analyses, false-positive and false-negative results can and do occur. Eurofins EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the data contained in, or any actions taken or omitted in reliance upon, this report.

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

Client: PVH Corp
C/O: Mr. Lou Schwartz
Re: JONESVILE ARU TESTING; CASSETTES

Date of Sampling: 01-19-2021
Date of Receipt: 01-20-2021
Date of Report: 01-21-2021

MoldSCORE™: Spore Trap Report

Outdoor Sample: 2 OUTSIDE

Fungi Identified	Outdoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3
Generally able to grow indoors*			
Alternaria		ND	< 13
Bipolaris/Drechslera group		ND	< 13
Chaetomium		ND	< 13
Cladosporium		ND	< 13
Curvularia		ND	< 13
Nigrospora		ND	< 13
Penicillium/Aspergillus types†		1	53
Stachybotrys		ND	< 13
Torula		ND	< 13
Seldom found growing indoors**			
Ascospores		ND	< 13
Basidiospores		9	480
Rusts		ND	< 13
Smuts, Periconia, Myxomycetes		1	13
Total			**547**

Location: 1 ARU #9

Fungi Identified	Indoor sample spores/m3 <100 1K 10K >100K	Raw count	Spores/ m3	MoldSCORE‡ 100 200 300	Score
Generally able to grow indoors*					
Alternaria		ND	< 13		100
Bipolaris/Drechslera group		ND	< 13		100
Chaetomium		ND	< 13		100
Cladosporium		ND	< 13		100
Curvularia		ND	< 13		100
Nigrospora		ND	< 13		100
Penicillium/Aspergillus types†		1	53		108
Stachybotrys		ND	< 13		100
Torula		ND	< 13		100
Seldom found growing indoors**					
Ascospores		ND	< 13		100
Basidiospores		1	53		103
Rusts		ND	< 13		100
Smuts, Periconia, Myxomycetes		ND	< 13		100
Total			**107**	**Final MoldSCORE**	**108**

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

Client: PVH Corp
C/O: Mr. Lou Schwartz
Re: JONESVILE ARU TESTING; CASSETTES

Date of Sampling: 01-19-2021
Date of Receipt: 01-20-2021
Date of Report: 01-21-2021

MoldSCORE™: Spore Trap Report

* The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

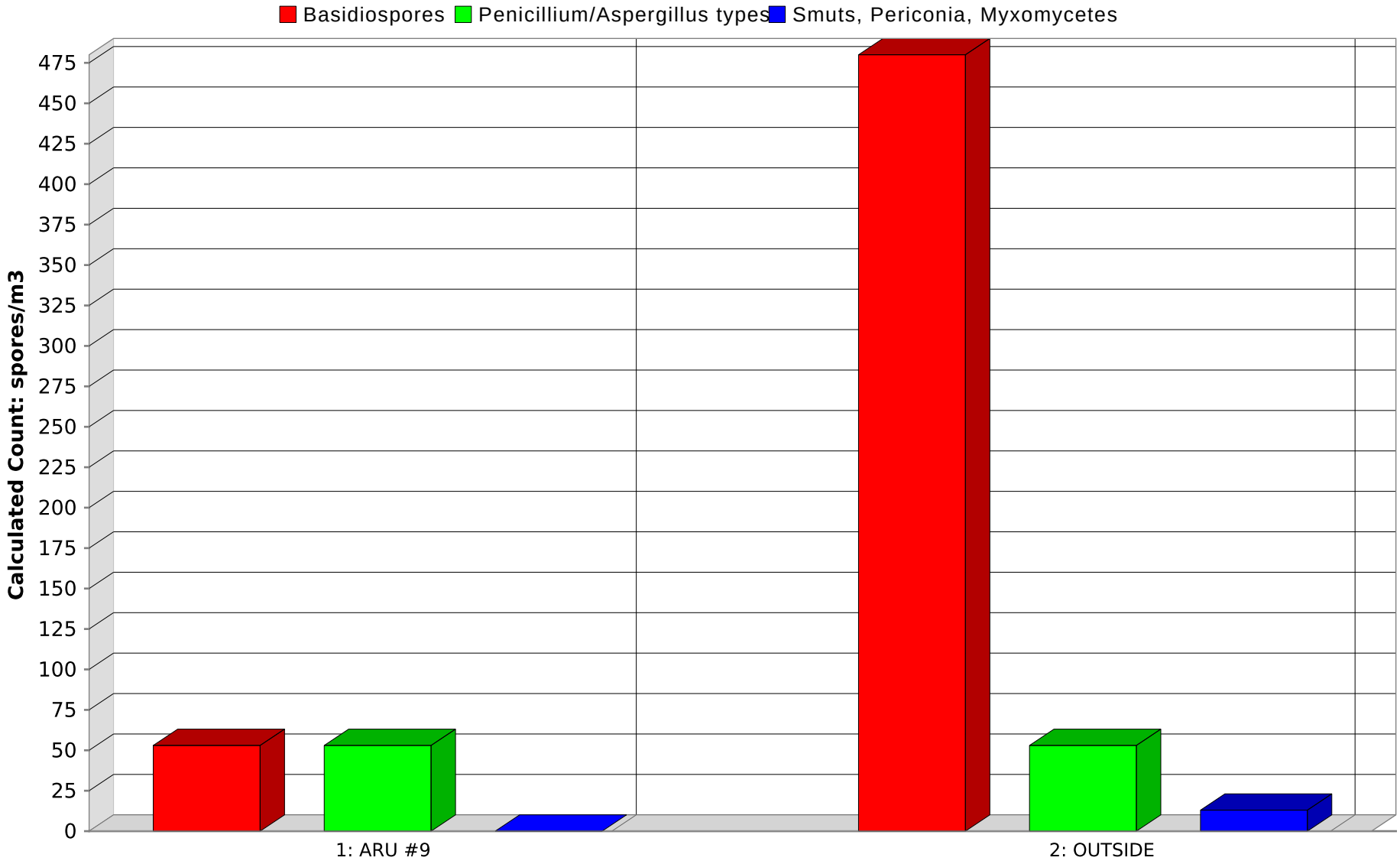
** These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

†The spores of Aspergillus and Penicillium (and others such as Acremonium, Paecilomyces) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods.

‡Rated on a scale from 100 to 300. A rating less than 150 is low and indicates a low probability of spores originating inside. A rating greater than 250 is high and indicates a high probability that the spores originated from inside, presumably from indoor mold growth. A rating between 150 and 250 indicates a moderate likelihood of indoor fungal growth. MoldSCORE is NOT intended for wall cavity samples. It is intended for ambient air samples in residences. Using the analysis on other samples (like wall cavity samples) will lead to misleading results.

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

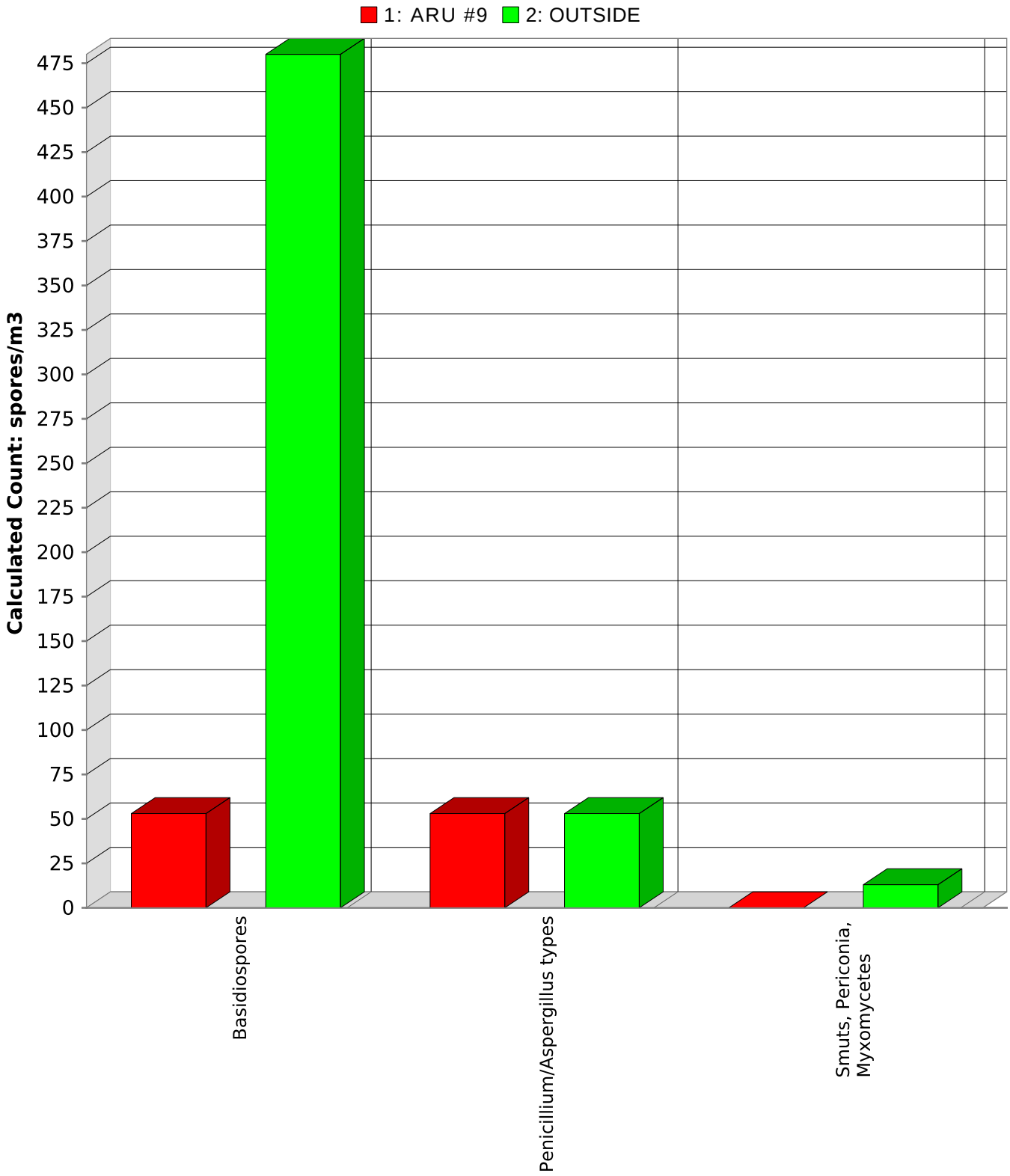


Comments:

Note: Graphical output may understate the importance of certain "marker" genera.

Eurofins EMLab P&K
3929 Old Lee Highway, Suite 91C, Fairfax, VA 22030
(866) 871-1984 Fax (856) 334-1040 www.emlab.com

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY



Comments:

Note: Graphical output may understate the importance of certain "marker" genera.

MICROBAC®

Volume _____

FINAL REPORT

VIRUCIDAL HARD-SURFACE EFFICACY TEST – Severe Acute Respiratory Syndrome-related Coronavirus 2 (SARS-CoV-2) (COVID-19 Virus)

Test Substance
PATH-AWAY ANTI-PATHOGENIC AEROSOL SOLUTION

Lot Numbers
22020
42020
52020

Test Organism
Severe Acute Respiratory Syndrome-related Coronavirus 2 (SARS-CoV-2) (COVID-19 Virus),
Strain: USA-WA1/2020, Source: BEI Resources, NR-52281

Test Guidelines
EPA (2018) Guidelines 810.2000 and 810.2200 (G)

Author
Cameron Wilde

Study Completion Date
10/12/20

Performing Laboratory
Microbac Laboratories, Inc.
105 Carpenter Drive
Sterling, VA 20164

Laboratory Project Identification Number
1029-102

Protocol Identification Number
GLO.1.07.01.20

Sponsor
Global Infection Control Consultants, LLC
23 Countryside Court
Bluffton, SC 29909

STATEMENT OF NO DATA CONFIDENTIALITY

No claim of confidentiality, on any basis whatsoever, is made for any information contained in this document. I acknowledge that information not designated as within the scope of FIFRA sec.10(d)(1)(A), (B) or (C) and which pertains to a registered or previously registered pesticide is not entitled to confidential treatment and may be released to the public, subject to the provisions regarding disclosure to multinational entities under FIFRA 10(g).

Submitter signature: _____ Date:_____
Printed Name of Signer: _____
Printed Name of Company: _____

GOOD LABORATORY PRACTICE COMPLIANCE STATEMENT

The following is a detailed description of all differences between the practices used in the study and those required by 40 CFR part § 160:

- Information on the identity, strength, purity, stability, uniformity, and dose solution analysis of the test substance resides with the sponsor of the study.

Study Director Signature: _____ Date: 10/12/2020
Typed Name: Cameron J. Wilde
Typed Name of Laboratory: Microbac Laboratories, Inc.

Sponsor Signature: _____ Date: _____
Printed Name: _____
Printed Name of Company: _____

Submitter Signature: _____ Date: _____
Printed Name: _____
Printed Name of Company: _____

QUALITY ASSURANCE UNIT STATEMENT

The Quality Assurance Unit of Microbac has inspected Project Number 1029-102 to be in compliance with current Good Laboratory Practice regulations (40 CFR § 160).

The dates that inspections were made and the dates that findings were reported to management and to the study director are listed below.

Phase Inspected	Date of Inspection	Date Reported to Study Director	Date Reported to Management
Protocol	08/14/20	08/14/20	08/14/20
In Process (Incubation)	08/14/20	08/14/20	08/14/20
Final Report	10/08/20	10/08/20	10/08/20

Lucas Thurn, RQAP-GLP
Quality Assurance Associate III

10/12/2020
Date

TABLE OF CONTENTS

Final Report – Cover Page 1

Statement of No Data Confidentiality 2

Good Laboratory Practice Compliance Statement 3

Quality Assurance Unit Statement 4

Table of Contents 5

Test Substance Characterization 6

Test Summary 6-7

Test Procedures 8-10

Protocol Changes 11

Study Dates and Facilities 11

Records to be Maintained 11

Test Acceptance Criteria 11

Calculations 12

Results 12-15

Test Substance Evaluation Criteria 16

Conclusions 16

References 17

Appendix I 18

Signed Protocol 19-33

Project Sheet(s) 34

Appendix II 35

Certificate(s) of Analysis 36-38

TEST SUBSTANCE CHARACTERIZATION

Test Substance characterization as to the identity, strength, purity, solubility and composition, as applicable, according to 40 CFR, Part 160, Subpart F [160.105] was documented prior to its use in the study. The Test Substance Certificate of Analysis Reports, provided by the sponsor, are found in Appendix II.

TEST SUMMARY

Study Title:	VIRUCIDAL HARD-SURFACE EFFICACY TEST – Severe Acute Respiratory Syndrome-related Coronavirus 2 (SARS-CoV-2) (COVID-19 Virus)
Project No.:	1029-102
Protocol No.:	GLO.1.07.01.20
Test Method:	ASTM International E1053-20 “Standard Test Method to Assess Virucidal Activity of Chemicals Intended for Disinfection of Inanimate, Nonporous Environmental Surfaces”
Sponsor:	Global Infection Control Consultants, LLC 23 Countryside Court Bluffton, SC 29909
Testing Facility:	Microbac Laboratories, Inc. 105 Carpenter Drive Sterling, VA 20164
Study Objective:	This test was performed in order to substantiate virucidal efficacy claims for a test substance by determining the efficacy of the test substance to disinfect hard surfaces contaminated with SARS-CoV-2. This test was designed to simulate consumer use and was performed in conformance to EPA OCSPP 810.2000 and 810.2200 Product Performance Test Guidelines.
Study Dates:	Study Initiation: 08/13/20 Experimental Start: 08/13/20 Experimental End: 08/20/20 Study Completion: See page 1

TEST SUMMARY (continued)

Test Substance: PATH-AWAY ANTI-PATHOGENIC AEROSOL SOLUTION

- Lot No.: 22020, Received: 06/26/20, assigned DS No. K889
- Lot No.: 42020, Received: 06/26/20, assigned DS No. K890
- Lot No.: 52020, Received: 06/26/20, assigned DS No. K891
- Physical Description: Liquid
- Storage Condition: Dark, Ambient Room Temperature
- Active Ingredients: Citrus Extract, Ascorbic Acid, Glycerine
- Dilution: Ready-to-use
- Diluent: Not applicable

Test Conditions:

Organic Soil Load:	5.0% FBS in viral inoculum
Contact Time:	2 minutes and 5 minutes
Contact Temperature:	21°C
Contact Relative Humidity:	53%

Challenge Virus: Severe Acute Respiratory Syndrome-related Coronavirus 2 (SARS-CoV-2) (COVID-19 Virus)

- Strain: USA-WA1/2020
- Source: BEI Resources, NR-52281

Indicator Cells: Vero E6 cells

- Source: ATCC CRL-1586

Other Reagents: Not applicable

Incubation Time: 7 days

Incubation Temperature: 36 ± 2°C with 5 ± 3% CO_2

Dilution Medium (DM): Minimum Essential Medium (MEM) + 2% Newborn Calf Serum (NCS)

Neutralizer: MEM + 10% NCS + 2% HEPES + 0.5% Polysorbate-80 + 0.01 N NaOH

Study Design: This study was performed according to the signed protocol and project sheet(s) issued by the Study Director (see Appendix I).

Study Personnel:

Cameron J. Wilde	Senior Scientist (Study Director
Brandon G. Narvaez	Associate Scientist II

TEST PROCEDURES

Indicator Cells:

Vero E6 cells were obtained from ATCC and maintained in cell culture at 36 ± 2°C with 5 ± 3% CO_2 prior to seeding. The indicator cell plates were prepared 12 – 30 hours prior to inoculation with test sample. The cells were seeded in 24-well plates at a density of 1.5 x 10^5 cells/mL at 1 mL per well.

Virus Inoculum:

The original stock virus was suspended in MEM + 5% FBS, aliquoted, and stored at -60 to -90°C. Frozen viral stock was thawed on the day of the test.

Challenge Virus:

Virus was not diluted and contained a 5.0% FBS load.

Test Substance:

The test substance was received ready-to-use. The test substance did not require equilibration to the contact temperature prior to use as it was stored at ambient room temperature.

Test Carriers:

Glass carriers were inoculated with 0.4 mL of virus inoculum and dried for 50 minutes at 21°C with 53% Relative Humidity (RH).

Test Substance Application and Exposure Conditions:

2.0 mL of test substance was added to the dried virus inoculum and held for the contact time of 2 minutes and 5 minutes at 21°C with 53% RH.

TEST PROCEDURES (continued)

Recovery of Samples:

After each contact time, the test substance was neutralized with 2.0 mL of neutralizer. The mixture was scraped from the surface of the carrier with a cell scraper. This post-neutralized sample (PNS) was considered the 10^{-1} dilution. An aliquot of the PNS was ten-fold serially diluted in DM.

Infectivity Assay:

Selected dilutions of the sample were inoculated onto the plates at 1.0 mL per well, 4 wells per dilution, and incubated at 36 ± 2°C with 5 ± 3% CO_2. After 7 days, the plates were removed from incubation, scored, and recorded for test-substance specific cytotoxic effects and/or virus-specific cytopathic effect (CPE).

Neutralizer Effectiveness and Viral Interference Control (NE/VI):

The control was performed using the longer contact time to assess whether residual active ingredient was present after neutralization (Neutralizer Effectiveness) or if the neutralized test substance interferes with virus infectivity (Viral Interference). The NE/VI was prepared identically to the test sample except DM was used in lieu of virus inoculum to inoculate the carrier. After test substance application and neutralization, the PNS was divided into two portions, one for the NE/VI and one for the Cytotoxicity (see below). For the NE/VI, a 0.5 mL aliquot of the PNS was ten-fold serially diluted and 100 µL of virus stock (containing 1000 $TCID_{50}$ units per well) was added individually to selected dilutions and held for at least the contact time. Selected dilutions were inoculated onto indicator cell plates and incubated in an identical manner as the test samples.

Cytotoxicity Control (CT):

This control was performed using the longer contact time to assess the cytotoxic effects of the test substance on indicator cells. The CT (obtained from the NE/VI) was prepared identically to the NE/VI except no virus was added to the selected dilutions inoculated onto indicator cells plates and incubated in an identical manner as the test samples.

Plate Recovery Control (PRC):

This control was performed using the longer contact time to establish the input viral load to compare with the test substance results to evaluate the viral reduction by the test substance. The PRC was prepared identically to the test sample except DM was used in lieu of test substance to treat the dried virus inoculum during test substance application. Selected dilutions were inoculated onto indicator cell plates and incubated in an identical manner as the test samples.

TEST PROCEDURES (continued)

Cell Viability Control (CVC):

This control was performed to demonstrate that the indicator host cells remained viable and to confirm the sterility of the media employed throughout the incubation period. Indicator cell plates were aspirated, and 1.0 mL of DM was added to 4 wells of indicator cells and incubated in an identical manner as the test samples.

Virus Stock Titer Control (VST):

This control was performed to demonstrate that the titer of the stock virus was appropriate for use and that the viral infectivity assay was performed appropriately. An aliquot of the virus inoculum used in the study was ten-fold serially diluted in DM. Selected dilutions were inoculated onto indicator cell plates and incubated in an identical manner as the test samples.

PROTOCOL CHANGES

Protocol Amendments:

No protocol amendments occurred during this study.

Protocol Deviations:

No protocol deviations occurred during this study.

STUDY DATES AND FACILITIES

The laboratory phase of this test was performed at Microbac Laboratories, Inc., 105 Carpenter Drive, Sterling, VA 20164, from 08/13/2020 – 08/20/2020. The study director signed the protocol on 08/13/2020. The study completion date is the date the study director signed the final report. The individual test dates are as follows:

- Testing started at 4:29 pm on 08/13/2020 and ended at 5:10 pm on 08/20/2020.

All changes or revisions of the protocol were documented, signed by the study director, dated and maintained with the protocol.

RECORDS TO BE MAINTAINED

All testing data, protocol, protocol modifications, test substance records, the final report, and correspondence between Microbac and the sponsor will be stored in the archives at Microbac Laboratories, Inc., 105 Carpenter Drive, Sterling, VA 20164, or at a controlled facility off site.

TEST ACCEPTANCE CRITERIA

The test was considered acceptable for test substance evaluation due to the criteria below being satisfied:

- The infectious virus recovered from the PRC was ≥ 4.8 Log_{10} $TCID_{50}$ units.
- Viral-induced CPE was distinguishable from test substance induced cytotoxicity (if any).
- Virus was recovered from dilutions of the NE/VI control not exhibiting cytotoxicity.
- The CVC did not exhibit CPE.

CALCULATIONS

Titer Calculation:

The 50% Tissue Culture Infectious Dose per mL ($TCID_{50}$/mL) was determined using the Spearman-Karber method using the following formula:

$$m = x_k + \left(\frac{d}{2}\right) - d\sum p_i$$

where: m = the logarithm of the dilution at which half of the wells are infected relative to the test volume
x_k = the logarithm of the smallest dosage which induces infection in all cultures
d = the logarithm of the dilution factor
p_i = the proportion of positive results at dilution i
$\sum p_i$ = the sum of p_i (starting with the highest dilution producing 100% infection)

The values were converted to $TCID_{50}$/mL using a sample inoculum of 1.0 mL.

Viral Load Calculation:

Load (Log_{10} $TCID_{50}$) per carrier = Titer (Log_{10} $TCID_{50}$/mL) + Log_{10} [volume per sample (mL)]

Viral Reduction Calculation:

Log_{10} Reduction = Initial Viral Load (Log_{10} $TCID_{50}$*) – Output Viral Load (Log_{10} $TCID_{50}$*)

***** per assayed volume and per carrier

RESULTS

Results are presented in Tables 1 – 6.

Key (for all tables):

T/y = Cytotoxicity observed in y wells inoculated; viral cytopathic effects (CPE) could not be determined
X/y = X wells out of y wells inoculated exhibited positive viral cytopathic effect
0/y = 0 out of y wells inoculated exhibited positive viral CPE; no cytotoxicity or bacterial contamination was observed in any of the wells inoculated

RESULTS (continued)

Table 1
Plate Recovery Control (PRC)

Dilution*	PRC
	Replicate 1
10^{-3}	4/4
10^{-4}	4/4
10^{-5}	4/4
10^{-6}	4/4
10^{-7}	2/4
10^{-8}	0/4
Titer (Log_{10} $TCID_{50}$/mL)	7.00
Load (Log_{10} $TCID_{50}$)**	6.60

*Dilution refers to the fold of dilution from the virus inoculum.
**Per carrier (0.40 mL of Undilute [10^0])

Table 2
Test Substance

Dilution*	Path-Away Anti-Pathogenic Aerosol Solution		
	2 minutes		
	Lot No. 22020	Lot No. 42020	Lot No. 52020
10^{-2}	T/4	T/4	T/4
10^{-3}	T/4	T/4	T/4
10^{-4}	0/4	0/4	0/4
10^{-5}	0/4	0/4	0/4
10^{-6}	0/4	0/4	0/4
10^{-7}	0/4	0/4	0/4
Titer (Log_{10} $TCID_{50}$/mL)	≤ 3.50	≤ 3.50	≤ 3.50
Load (Log_{10} $TCID_{50}$)**	≤ 3.10	≤ 3.10	≤ 3.10
Log_{10} Reduction***	≥ 3.50	≥ 3.50	≥ 3.50

*Dilution refers to the fold of dilution from the virus inoculum.
**Per carrier (0.40 mL of Undilute [10^0])
***Per assayed volume and per carrier

RESULTS (continued)

Table 3
Test Substance

Dilution*	Path-Away Anti-Pathogenic Aerosol Solution		
	5 minutes		
	Lot No. 22020	Lot No. 42020	Lot No. 52020
10^{-2}	T/4	T/4	T/4
10^{-3}	T/4	T/4	T/4
10^{-4}	0/4	0/4	0/4
10^{-5}	0/4	0/4	0/4
10^{-6}	0/4	0/4	0/4
10^{-7}	0/4	0/4	0/4
Titer (Log_{10} $TCID_{50}$/mL)	≤ 3.50	≤ 3.50	≤ 3.50
Load (Log_{10} $TCID_{50}$)**	≤ 3.10	≤ 3.10	≤ 3.10
Log_{10} Reduction***	≥ 3.50	≥ 3.50	≥ 3.50

*Dilution refers to the fold of dilution from the virus inoculum.
**Per carrier (0.40 mL of Undilute [10^0])
***Per assayed volume and per carrier

Table 4
Neutralizer Effectiveness/Viral Interference (NE/VI) and Cytotoxicity (CT) Controls

Dilution*	Path-Away Anti-Pathogenic Aerosol Solution					
	Lot No. 22020		Lot No. 42020		Lot No. 52020	
	NE/VI	CT	NE/VI	CT	NE/VI	CT
10^{-2}	T/4	T/4	T/4	T/4	T/4	T/4
10^{-3}	T/4	T/4	T/4	T/4	T/4	T/4
10^{-4}	4/4	0/4	4/4	0/4	4/4	0/4

*Dilution refers to the fold of dilution from the mock inoculum.

RESULTS (continued)

Table 5
Cell Viability Control (CVC)

CVC
0/4
Cells were viable; media was sterile

Table 6
Virus Stock Titer Control (VST)

Dilution*	VST
10^{-4}	4/4
10^{-5}	4/4
10^{-6}	4/4
10^{-7}	4/4
10^{-8}	1/4
10^{-9}	0/4
Titer (Log_{10} $TCID_{50}$/mL)	7.75

*Dilution refers to the fold of dilution from the virus inoculum.

TEST SUBSTANCE EVALUATION CRITERIA

According to the US Environmental Protection Agency, the test substance passes the test if the following criteria are met:

- The test substance must demonstrate a ≥ 3 Log_{10} reduction on each test carrier in the presence or absence of cytotoxicity. If cytotoxicity is present, the virus control titer should be sufficient to demonstrate a ≥ 3 Log_{10} reduction in viral titer on each test carrier beyond the level of cytotoxicity.

CONCLUSIONS

When tested as described, PATH-AWAY ANTI-PATHOGENIC AEROSOL SOLUTION, Lot Nos. 22020, 42020, and 52020, passed the Virucidal Hard-Surface Efficacy Test when Severe Acute Respiratory Syndrome-related Coronavirus 2 (SARS-CoV-2) (COVID-19 Virus), containing 5.0% FBS, was exposed to the test substance for 2 minutes and 5 minutes at 21°C and 53% RH.

All controls met the criteria for a valid test. These conclusions are based on observed data.

REFERENCES

1. ASTM E1053-20, Standard Test Method to Assess Virucidal Activity of Chemicals Intended for Disinfection of Inanimate, Nonporous Environmental Surfaces, ASTM International, West Conshohocken, PA, 2011.

2. U.S. Environmental Protection Agency, Office of Chemical Safety and Pollution Prevention, Product Performance Test Guidelines, OCSPP 810.2200: Disinfectants for Use on Environmental Surfaces, Guidance for Efficacy Testing, February 2018.

3. U.S. Environmental Protection Agency, Office of Chemical Safety and Pollution Prevention, Product Performance Test Guidelines, OCSPP 810.2000: General Considerations for Testing Public Health Antimicrobial Pesticides, Guidance for Efficacy Testing, February 2018.

4. U.S. Environmental Protection Agency, Office of Chemical Safety and Pollution Prevention, Product Performance Test Guidelines, Frequently Asked Questions (FAQ) for OCSPP 810.2000, 810.2100, 810.2200.

APPENDIX I

MICROBAC

Microbac Protocol

VIRUCIDAL HARD-SURFACE EFFICACY TEST -

Severe Acute Respiratory Syndrome-related Coronavirus 2 (SARS-CoV-2) (COVID-19 Virus)

Testing Facility
Microbac Laboratories, Inc.
105 Carpenter Drive
Sterling, VA 20164

Prepared for
Global Infection Control Consultants LLC
23 Countryside Court
Bluffton, SC 29909

July 1, 2020

Microbac Protocol: GLO.1.07.01.20

Microbac Project: 1029-102

AVM

OBJECTIVE:

This test is designed to substantiate virucidal effectiveness claims for a test substance to be labeled as a virucide. It determines the potential of the test substance to disinfect hard surfaces contaminated with the test virus. The test is designed to simulate consumer use and conforms to EPA OCSPP 810.2000 (2018) and 810.2200 (2018) Product Performance Test Guidelines, Frequently Asked Questions (FAQ) for OCSPP 810.2000, 810.2100, and 810.2200, and follows the procedure outlined in the ASTM International test method designated E1053-20, "Standard Test Method to Assess Virucidal Activity of Chemicals Intended for Disinfection of Inanimate, Nonporous Environmental Surfaces".

TESTING CONDITIONS:

Virus will be dried on a suitable sterile hard surface at ambient temperature. One test substance (liquid), three batches (lots), will be tested at two contact times and one replicate (N=1). The test substance will be used to treat the dried virus on a glass Petri dish carrier. After a defined exposure period as specified by the sponsor, the test substance-virus mixture will be neutralized, scraped off from the surface, collected, and tested for the presence of infectious virions.

MATERIALS:

A. Test, control and reference substances will be supplied by the Sponsor of the study. Microbac will append the Sponsor-provided Certificate(s) of Analysis (CoA) to this study report, as per CFR 40.160.105:

- The identity, strength, purity, and composition, or other characteristics which will appropriately define the test, control, or reference substance shall be determined and shall be documented by the sponsor before its use in a study. Methods of synthesis, fabrication, or derivation of the test, control, or reference substance shall be documented and retained by the sponsor.

- When relevant to the conduct of the study the solubility of each test, control, or reference substance shall be determined by the sponsor before the experimental start date. The stability of the test, control, or reference substance shall be determined by the sponsor before the experimental start date or concomitantly according to written standard operating procedures, which provide for periodic analysis.

The test substance will be tested as supplied by the sponsor unless directed otherwise. All operations performed on the test substance such as dilution or specialized storage conditions must be specified by the sponsor before initiation of testing.

The sponsor assures Microbac testing facility management that the test substance has been appropriately tested for identity, strength, purity, stability, and uniformity as applicable.

Microbac will retain all unused test substances for a period of one year upon completion of the test, and then discard them in a manner that meets the approval of the safety officer or return them to the Sponsor. The test materials and the paper records will be retained in accordance to FIFRA. Microbac will contact the Study Sponsor to arrange for transfer of records when/if the test substance is returned to the Sponsor.

B. Materials supplied by Microbac, including, but not limited to:

1. Challenge virus (requested by the sponsor of the study): Severe Acute Respiratory Syndrome-related Coronavirus 2 (SARS-CoV-2) (COVID-19 Virus), Strain: USA-WA1/2020, Source: BEI Resources, NR-52281

2. Host cell line: Vero E6 cells, ATCC CRL-1586

3. Laboratory equipment and supplies.

4. Media and reagents:

   Media and reagents relevant to the virus-host system and test substance being tested will be documented in the first project sheet and data pack.

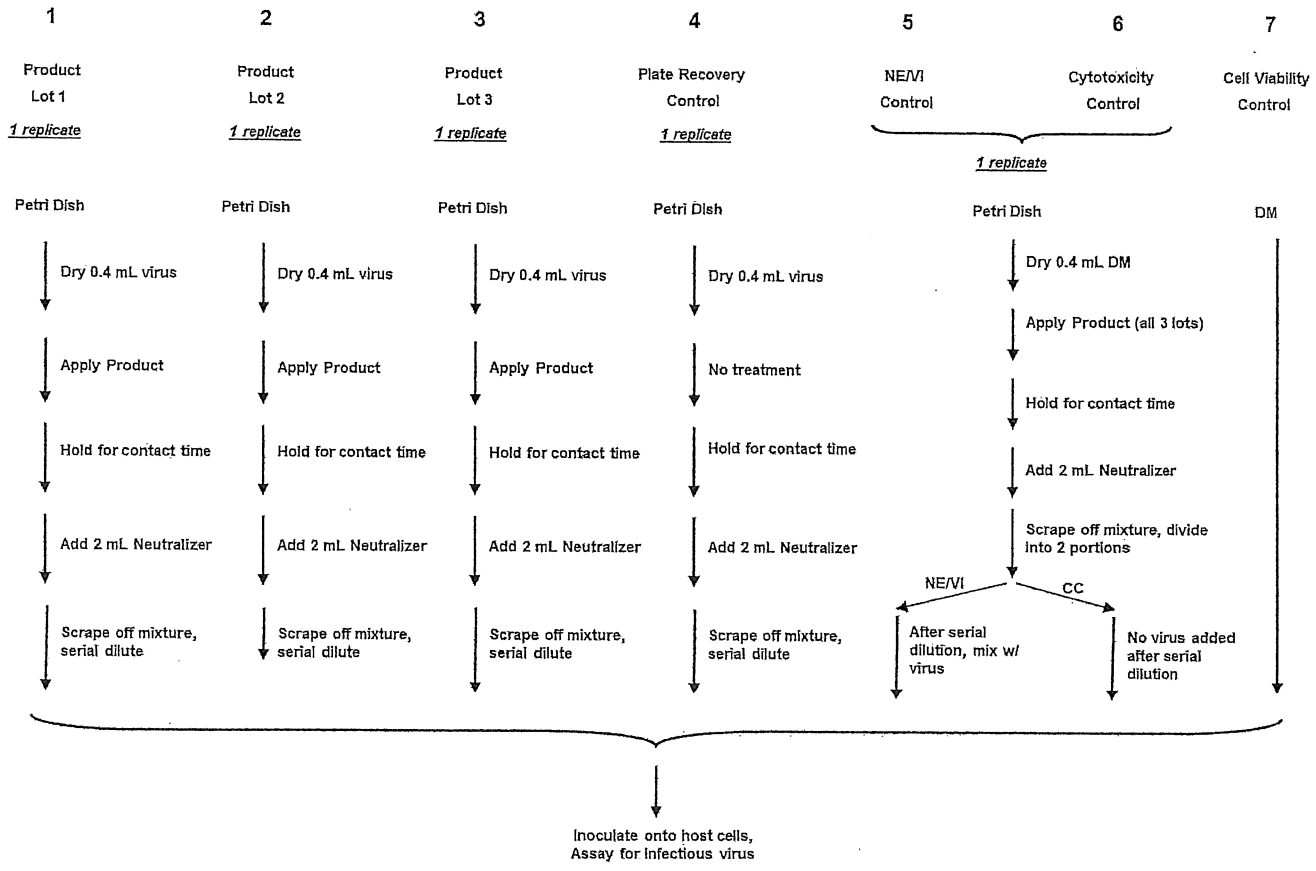
TEST SYSTEM IDENTIFICATION:

All Petri dishes, dilution tube racks, and host-containing apparatus will be appropriately labeled with the following information: virus, host, and test substance and/or project number.

EXPERIMENTAL DESIGN:

All of the procedures involved in performance of this study are described in a detailed series of SOPs that are maintained at Microbac. SOPs and Logs are referred to in the raw data and are required as part of GLP regulations. The study flow diagram is shown in Figure 1, with details described in the following sections.

FIGURE 1



DM: Dilution Medium
NE/VI: Neutralizer Effectiveness/Viral Interference control
CT: Cytotoxicity Control

Note: One test substance, three lots, will be tested at two exposure (contact) times and one replicate (N=1). The NE/VI and CT controls will be performed at one replicate per lot.

A. Inoculum preparation:

Viral stocks are purchased from reputable sources that identify them by scientifically accepted methods and may have been propagated at Microbac. Records are maintained that demonstrate the origin of the virus. The virus stocks are stored at an ultra-low temperature.

Frozen viral stocks will be thawed on the day of the test. Serum will be added to viral stock to achieve an organic load of 5.0% (if not already 5.0%), unless otherwise directed by the Sponsor and pre-agreed by Microbac. If the challenge virus culture is standardized by concentration or dilution, or if a column is used, these manipulations must be documented and reported.

Note: a level of approximately 4.8 – 6.8 Log_{10} virus challenge (as indicated by the plate recovery control load) when there is no cytotoxicity associated with the test substance, or approximately 3.0 – 5.0 Log_{10} beyond the level of cytotoxicity when present, should be achieved whenever possible.

B. Carrier preparation:

For each lot of the test substance, an aliquot of 0.4 mL of stock virus will be spread over the bottom of pre-sterilized glass Petri dishes. This volume will remain consistent among all test and control runs. Then the virus will be allowed to dry at ambient temperature. The drying time, temperature, and relative humidity will be recorded and reported.

Two carriers will be prepared for each lot of the test substance using virus. One carrier will be prepared for the plate recovery control using virus. Additionally, one carrier will be prepared for each lot of test substance for the neutralizer effectiveness/viral interference and cytotoxicity controls using media in lieu of virus as the inoculum.

C. Test substance preparation:

Note: Information on the identity, strength, purity, stability, uniformity, and dose solution analysis of the test substance resides with the sponsor of the study.

The test substance will be prepared exactly according to the sponsor's directions (if provided). If the sponsor requests dilution of the test substance, the diluted test

substance will be used for testing within three hours of preparation. The prepared test substance, if not within the stipulated test temperature range, will be pre-equilibrated to the test temperature prior to use in the study as applicable.

D. Test:

Three lots of the test substance (liquid) will be tested at two contact times and one replicate (N=1). Note: The temperature and relative humidity during the exposure period will be recorded and reported.

For direct liquid application test substance, for each run, after the inoculum has dried, 2.0 mL of the test substance will be added. The dried virus film must be completely covered by the test substance. The plates will remain at the temperature and for the time specified by the sponsor. After the contact period, the test agent will be neutralized with 2.0 mL of appropriate neutralizer and the mixture will be scraped from the surface of the dish with a cell scraper. This post-neutralized sample (PNS) will be considered approximately a 10^{-1} dilution.

For spray type test substance, an aliquot of the test substance, ready-to-use, will be dispensed into a sterilized spray bottle. The spray bottle will then be shaken 2 – 3 times to ensure homogeneity and sprayed to charge the spray bottle. A mock spray action will be performed by applying the test substance as the sponsor directs onto at least two blank Petri dishes. Then the volume dispensed onto each dish will be measured and averaged. This averaged volume from the mock spray runs will be used for the neutralizer for all applicable runs and for the Plate recovery control runs. Then the test substance will be sprayed onto the virus carriers in a horizontal position until thoroughly wet from a distance of 6" – 8". Each carrier will be held in a horizontal position for the exposure time as specified by the sponsor. After the contact period, the test substance will be neutralized with an appropriate neutralizer using the averaged volume from the mock spray runs; and the mixture will be scraped off from the surface of the dish with a cell scraper. This post-neutralized sample (PNS) will be considered approximately a 10^{-1} dilution.

If Sephacryl columns are used to aid in the neutralization and to further reduce the cytotoxicity, each inoculum/test substance/neutralizer mixture sample will be loaded onto a pre-spun Sephacryl column. Following the passage through columns, the eluates will be aseptically collected and serially ten-fold diluted in DM. If columns are not used, serial ten-fold dilutions of the inoculum/test substance/neutralizer mixture will directly be prepared in DM.

E. Infectivity assay:

The residual infectious virus in all test and control samples will be detected by viral-induced cytopathic effect (CPE).

Selected dilutions of the neutralized inoculum/test substance mixture (test samples) and control samples will be added to cultured host cells (at least four wells per dilution, per reaction mixture) and incubated at 36±2°C with 5±3% CO_2 for total 4 – 9 days. The host cells may be washed twice with phosphate buffered saline prior to inoculation. The inoculated culture will be observed and refed with fresh media as necessary, during the incubation period. These activities, if applicable, will be recorded. The host cells will then be examined microscopically for presence of infectious virions. The resulting virus-specific CPE and test substance-specific cytotoxic effects will be scored by examining all test and control samples. These observations will be recorded.

F. Controls:

1. Plate recovery control (PRC):

This control will be performed in a single run, concurrently with the test substance runs using the longest contact time as worst case.

The virus inoculum will be spread over the surface of a sterile glass Petri dish and left to dry at ambient temperature. A volume of DM equivalent to that of the test substance will be added to the dried virus. Post-contact time, virus will be subjected to the identical neutralization procedure as the test substance. This control will determine the relative loss in virus infectivity resulting from drying and neutralization alone.

The results from this control will be compared with the test results to confirm recovery of at least 4.8-Log_{10} per carrier of infectious virus in this control following drying and neutralization. Its titer will be used to compare with the titers of the test results to reach the acceptable test criteria (see below).

2. Neutralizer effectiveness/Viral interference control (NE/VI):

This control will determine if residual active ingredient is present after neutralization and if the neutralized test substance interferes with the virus

infection system. This control will be performed for each lot of test substance at one replicate using the longest contact time as worst case.

The test substance will be processed exactly as the test procedure but in lieu of virus inoculum, dried DM will be exposed to the test substance and assayed as previously described. Post-treatment and neutralization, the neutralized DM/test substance mixture will be divided into two portions, one for cytotoxicity control and the other for neutralizer effectiveness/viral interference control and processed as the test.

If columns are used, each portion will be passed through individual columns and the eluate will be serially diluted ten-fold in DM. If columns are not used, each portion will be directly diluted using serial ten-fold dilutions in DM.

The neutralizer effectiveness/viral interference control sample will be diluted as follows: using dilution test tubes and appropriate pipette, an aliquot of the PNS will be used for making serial 10-fold dilutions in DM (for example, 0.5 mL sample + 4.5 mL DM). Following serial dilution, 0.1 mL of a low titered virus, containing approximately 1,000 – 5,000 infectious units of virus, will be added to 4.5 mL of each dilution and held for a period of no shorter than the contact time. Then these samples will be used to inoculate host cells as described for the test procedure.

Selected dilutions of the sample will be added to cultured cell monolayers at a minimum of four wells per dilution per sample, as described in the "Infectivity Assay" section.

3. Cytotoxicity control (CT):

This control will be performed for each lot of test substance at one replicate.

The cytotoxicity sample, acquired from the neutralizer effectiveness/viral interference control run, will be diluted and have no virus added. Selected dilutions will be inoculated and incubated in the same manner as the rest of the test and control samples. These effects are distinct from virus-induced cytopathic effects, which will be evident in the plate recovery control cultures.

4. Column titer control (to be performed only if a Sephacryl column is used):

   This control will be performed to determine any affect the columns may have on infectious virus titer. It will be performed in a single run.

   The sample for this control will be acquired from a portion of the PRC, prior to passing through the columns and will be serially diluted in DM, then processed in the same manner as the test.

5. Cell viability control:

   This control will be performed in a single run. It will demonstrate that cells remain viable throughout the course of the assay period. In addition, it will confirm the sterility of the DM employed throughout the assay period. At least four wells of cells will receive only DM and will be incubated and processed with both test and other controls. This will serve as the negative control.

6. Virus Stock Titer control (VST)

   This control will be performed in a single run. An aliquot of the virus used in the study will be directly serially diluted and inoculated onto the host cells to confirm the titer of the stock virus. This control will demonstrate that the titer of the stock virus is appropriate for use and that the viral infectivity assay is performed appropriately.

G. Calculation:

The 50% tissue culture infective dose per mL ($TCID_{50}$/mL) will be determined using the method of Spearman-Karber (Kärber G., Arch. Exp. Pathol. Pharmakol. 1931, 162: 480-483) or other appropriate methods such as Reed and Muench (Am. J. of Hyg. 1938, 27:493). The $TCID_{50}$/carrier, i.e., the viral load per carrier, will be calculated as follows. These analyses will be described in detail in the final report. The test results will be reported as reduction of the virus titer post treatment with the test substance expressed as log_{10}.

<u>The Virus Load ($TCID_{50}$/carrier) will be calculated in the following manner</u>:
Virus Load (Log_{10} $TCID_{50}$) = Virus Titer (Log_{10} $TCID_{50}$/mL) + Log_{10} [Volume per sample (mL)]

The Log_{10} Reduction Factor (LRF) will be calculated in the following manner:
Log_{10} Reduction Factor = Initial viral load (Log_{10} $TCID_{50}$, per assayed volume and per carrier) – Output viral load (Log_{10} $TCID_{50}$, per assayed volume and per carrier)

TEST ACCEPTANCE CRITERIA:

The test will be acceptable for evaluation of the test results if the criteria listed below are satisfied. The study director may consider other causes that may affect test reliability and acceptance.

- The infectious virus recovered from the PRC control must be ≥ 4.8-log_{10} $TCID_{50}$ units.
- Viral-induced cytopathic effect must be distinguishable from test substance induced cytotoxic effects (if any).
- Virus must be recovered from the neutralizer effectiveness/viral interference control (not exhibiting cytotoxicity).
- The Cell Viability Control (assay negative control) must not exhibit virus.

TEST SUBSTANCE EVALUATION CRITERIA:

According to the US Environmental Protection Agency, the test substance passes the test if the following are met:

- The product must demonstrate a ≥ 3 log_{10} reduction on each surface in the presence or absence of cytotoxicity; and
- If cytotoxicity is present, the virus control titer should be increased to demonstrate a ≥ 3 log_{10} reduction in viral titer on each surface beyond the cytotoxic level.

PERSONNEL AND TESTING FACILITIES:

A study director will be assigned prior to initiation of the test. Resumes are maintained and are available on request. This study will be conducted at Microbac Laboratories, Inc., 105 Carpenter Drive, Sterling, Virginia 20164.

REGULATORY COMPLIANCE AND QUALITY ASSURANCE (GLP studies only):

This study will be performed in compliance with the US Environmental Protection Agency's Good Laboratory Practices (GLP) regulations, 40 CFR 160 (note: information on the identity, strength, purity, stability, uniformity, and dose solution analysis of the test substance resides with the sponsor of the study unless otherwise stated).

The Quality Assurance Unit of Microbac will inspect the conduct of the study for GLP compliance. The dates of the inspections and the dates that findings are reported to the study management and study director will be included in the final report.

PROTOCOL AMENDMENTS AND DEVIATIONS:

Any protocol amendment(s) and protocol deviation(s) identified will be reported in project sheet(s) and included in the final report.

REPORT FORMAT:

This report will contain all items required by 40 CFR Part 160.185 and EPA 810.2000 and be in compliance with EPA PR Notice 2011-3. Microbac employs a standard report format for each test design. Each final report will provide at least the following information:

- Sponsor identification
- Test substance identification
- Type of assay and project number
- Study start and end time (clock time)
- Interpretation of results and conclusions
- Test results presented in tabular form
- Methods and evaluation criteria, if applicable
- Dates of study initiation and completion (GLP studies only)
- Signed Quality Assurance and Compliance Statements (GLP studies only)
- Certificate of Analysis (for GLP studies only; if provided by the Sponsor)
- List of personnel involved in the study

RECORDS TO BE MAINTAINED:

For all GLP studies, the original signed final report or an electronic copy will be sent to the Sponsor. The original signed final report, or a copy thereof, will be maintained in the study file. If requested, a draft report will be provided to the Sponsor for review prior to finalization of the report.

All raw data, protocol, protocol modifications, test substance records, the final report (or copy thereof), and correspondence between Microbac and the sponsor will be stored in the archives at Microbac Laboratories, Inc., 105 Carpenter Drive, Sterling, Virginia 20164 or in a controlled facility off site.

All changes or revisions to this approved protocol will be documented, signed by the study director, dated and maintained with this protocol. The sponsor will be notified of any change, resolution, and impact on the study as soon as practical.

The proposed experimental start and termination dates; additional information about the test substance; challenge virus and host cell line monolayers used and the type of neutralizers employed in the test will be addressed in a project sheet issued separately for each study. The date the study director signs the protocol will be the initiation date. All project sheets issued will be forwarded to the study sponsor for appropriate action.

REFERENCES

1. ASTM E1053-20, Standard Test Method to Assess Virucidal Activity of Chemicals Intended for Disinfection of Inanimate, Nonporous Environmental Surfaces, ASTM International, West Conshohocken, PA, 2020.

2. U.S. Environmental Protection Agency, Office of Chemical Safety and Pollution Prevention, Product Performance Test Guidelines, OCSPP 810.2200: Disinfectants for Use on Environmental Surfaces, Guidance for Efficacy Testing, February 2018.

3. U.S. Environmental Protection Agency, Office of Chemical Safety and Pollution Prevention, Product Performance Test Guidelines, OCSPP 810.2000: General Considerations for Testing Public Health Antimicrobial Pesticides, Guidance for Efficacy Testing, February 2018.

4. U.S. Environmental Protection Agency, Office of Chemical Safety and Pollution Prevention, Product Performance Test Guidelines, Frequently Asked Questions (FAQ) for OCSPP 810.2000, 810.2100, and 810.2200.

MISCELLANEOUS INFORMATION:

The following information is to be completed by the sponsor prior to initiation of the study (please check all applicable open boxes):

A. Test substance information:

Test substance name	PATH-AWAY ANTI-PATHOGENIC AEROSOL SOLUTION		
Test substance batch numbers	22020	42020	52020
Manufacture Date	FEB 5, 2020	APRIL 10, 2020	MAY 19, 2020
Expiration Date	FEB 5, 2025	APRIL 10, 2025	MAY 19, 2025
Active ingredient(s)	CITRUS EXTRACT CAS# 92346-89-9 ASCORBIC ACID CAS #50-81-7 GLYCERINE USP CAS# 56-81-5		
Test substance storage conditions	☑ Ambient ☐ Refrigerated ☐ Other:_____		
Level of active ingredients in testing	■ Lower Certified Limit (LCL)[1] ☐ At or below nominal		
MSDS provided	☐ Yes ☐ No	C of A provided	☐ Yes ☐ No
Dilution	☑ Ready to use DO NOT DILUTE ☐ _____ (_____ parts test substance + _____ parts diluent)		
Diluent	☑ Not applicable ☐ _____ ppm ±2.9% AOAC hard water ☐ Other:_____		
Contact time 1	2 MINUTES		
Contact time 2	5 MINUTES		
Contact temperature	■ Room Temperature (20±1°C) ☐ Other :_____		
Organic Load	■ 5.0% serum in viral inoculum ☐ Other: _____		
Test substance application	■ Apply directly to dried virus via pipetting ☐ Spray from 6-8 inches until thoroughly wet		
Study conduct	■ GLP ☐ Non-GLP		
Report submission	■ EPA ☐ Health Canada ☐ Other: _____		

[1] US EPA stipulates that 3 lots of test substance be tested at or below LCL for COVID-19

PROTOCOL APPROVAL BY SPONSOR:

Sponsor Signature: _____ Date: 6 JULY, 2020

Printed Name: ARTHUR V. MARTIN Ph. D.

PROTOCOL APPROVAL BY STUDY DIRECTOR (Microbac):

Study Director Signature: _____ Date: 08/13/2020

Printed Name: Cameron J. Wilde

Date Issued: 08/13/20 Project Sheet No. 1 Page No. 1 Laboratory Project Identification No. 1029-102

STUDY TITLE: VIRUCIDAL HARD-SURFACE EFFICACY TEST – Severe Acute Respiratory Syndrome-Related Coronavirus 2 (SARS-CoV-2) (COVID-19 Virus)	**STUDY DIRECTOR:** Cameron Wilde Signature 08/13/2020 Date		
TEST MATERIAL(S): PATH-AWAY ANTI-PATHOGENIC AEROSOL SOLUTION	**BATCH (LOT) NO.** 22020 42020 52020	**DATE RECEIVED:** 06/26/20 06/26/20 06/26/20	**DS NO.** K889 K890 K891
PERFORMING DEPARTMENT(S): Virology and Toxicology	**STORAGE CONDITIONS:** Location: I5 ■Dark ■Ambient Room Temperature ☐ Desiccator ☐Freezer ☐Refrigerator ☐ Other:		

PROTECTIVE PRECAUTION REQUIRED: MSDS ☐ Yes / ■ No

PHYSICAL DESCRIPTION: ☐ Solid ■ Liquid ☐ Aerosol ☐ Other:

PURPOSE: See attached protocol. **AUTHORIZATION:** See client signature.

PROPOSED EXPERIMENTAL START DATE: 08/13/20 **TERMINATION DATE:** 08/22/20

CONDUCT OF STUDY: ☐ FDA ■ EPA ☐ R&D ■ GLP ☐ GCP ☐ Other:

SPONSOR: Global Infection Control Consultants, LLC 23 Countryside Court Bluffton, SC 29909	**CONTACT PERSON:** Arthur V. Martin, Ph.D. amartin@gicclic.com

TEST CONDITIONS:

Challenge organism: SARS-CoV-2, Strain: USA-WA1/2020, Source: BEI Resources, NR-52281

Host cell line: Vero E6, Source: ATCC CRL-1586

Organic load: 5.0% serum in virus inoculum

Dilution medium: Minimum Essential Medium (MEM) + 2% Newborn Calf Serum (NCS)

Active ingredient(s): Citrus Extract, Ascorbic Acid, Glycerine

Dilution: Ready-to-use

Neutralizer: MEM + 10% NCS + 0.5% Polysorbate-80 + 2% HEPES + 0.01N NaOH

Contact time(s): 2 minutes and 5 minutes

Contact temperature: Room Temperature (20±1°C)

Incubation time: 4 – 9 days

Incubation temperature: 36±2°C with 5±3% CO_2

Test product application: Apply directly to dried virus via pipetting

APPENDIX II

GLOBAL
Infection Control Consultants LLC

Certificate of Analysis

Product: Path-Away Anti-Pathogenic Aerosol Solution® 3% Solution Final Mix

Product Description: A proprietary non-metallic, organic antimicrobial and antifungal compound.
Complies with Federal and FDA Regulations 21 CFR 182.3013 and CFR 184.1540
USA EPA Registration Exempt as per FIFRA 25(b)

Chemical Description

Active Ingredients	Specifications	Result
Proprietary Citrus Extract CAS #92346-89-9	1.00 – 2.00%	1.15%
Ascorbic Acid CAS #50-81-7	1.25 - 1.75%	1.47%
Glycerine USP CAS #56-81-5	1.00 – 1.50%	1.11%
Inert Ingredients		
Citrus pulp CAS #68514-76-1	0.001 – 0.050%	0.010%
Dextrose CAS#492-62-6	0.05 – 0.25%	0.15%
Moisture CAS #7732-18-5	96.0 – 97.25%	96.11%

Physical Properties

Description	Specifications	Result
Appearance	Light to moderate golden viscous liquid	Light to moderate golden viscous liquid
Gardner Color –*Orbeco-Hellige Comparator*	3 – 9	N/A
Specific Gravity – *Optima OPD-E*	1.10 – 1.30	N/A
pH (d25°) – *Fisher Accumet AB150*	1.50 – 3.00	N/A
Flash Point (°F) - *Rapid Flash Closed-Cup Tester*	270 - 300	N/A
Infrared IR – *Spectrum Two Perkin/Elmer*	Reference Spectra	PASS

Batch Date: 2-10-2020 Batch # 22020

Shipping Date: 6/25/2020
Shipped to: Microbac Laboratories

Shelf Life 5 – 7 years

Name Arthur V. Martin Ph.D. President

Authorized Signature / Date 6/24/2020

Do NOT store in steel or metal containers. Store in plastic or glass containers only.

GLOBAL
Infection Control Consultants LLC

Certificate of Analysis

Product: Path-Away Anti-Pathogenic Aerosol Solution® 3% Solution Final Mix

Product Description: A proprietary non-metallic, organic antimicrobial and antifungal compound.
Complies with Federal and FDA Regulations 21 CFR 182.3013 and CFR 184.1540
USA EPA Registration Exempt as per FIFRA 25(b)

Chemical Description

Active Ingredients	Specifications	Result
Proprietary Citrus Extract CAS #92346-89-9	1.00 – 2.00%	1.15%
Ascorbic Acid CAS #50-81-7	1.25 - 1.75%	1.465%
Glycerine USP CAS #56-81-5	1.00 – 1.50%	1.02%
Inert Ingredients		
Citrus pulp CAS #68514-76-1	0.001 – 0.050%	0.015%
Dextrose CAS#492-62-6	0.05 – 0.25%	0.15%
Moisture CAS #7732-18-5	96.0 – 97.25%	96.20%

Physical Properties

Description	Specifications	Result
Appearance	Light to moderate golden viscous liquid	Light to moderate golden viscous liquid
Gardner Color –*Orbeco-Hellige Comparator*	3 – 9	N/A
Specific Gravity – *Optima OPD-E*	1.10 – 1.30	N/A
pH (d25°) – *Fisher Accumet AB150*	1.50 – 3.00	N/A
Flash Point (°F) - *Rapid Flash Closed-Cup Tester*	270 - 300	N/A
Infrared IR – *Spectrum Two Perkin/Elmer*	Reference Spectra	PASS

Batch Date: 4-15-2020 Batch # 42020

Shipping Date: 6/25/2020
Shipped to: Microbac Laboratories

Shelf Life 5 – 7 years

Name Arthur V. Martin Ph.D. President

Authorized Signature / Date 6/24/2020

Do NOT store in steel or metal containers. Store in plastic or glass containers only.

GLOBAL
Infection Control Consultants LLC

Certificate of Analysis

Product: Path-Away Anti-Pathogenic Aerosol Solution® 3% Solution Final Mix

Product Description: A proprietary non-metallic, organic antimicrobial and antifungal compound.
Complies with Federal and FDA Regulations 21 CFR 182.3013 and CFR 184.1540
USA EPA Registration Exempt as per FIFRA 25(b)

Chemical Description

Active Ingredients	Specifications	Result
Proprietary Citrus Extract CAS #92346-89-9	1.00 – 2.00%	1.30%
Ascorbic Acid CAS #50-81-7	1.25 - 1.75%	1.40%
Glycerine USP CAS #56-81-5	1.00 – 1.50%	1.20%
Inert Ingredients		
Citrus pulp CAS #68514-76-1	0.001 – 0.050%	0.005%
Dextrose CAS#492-62-6	0.05 – 0.25%	0.10%
Moisture CAS #7732-18-5	96.0 – 97.25%	95.995%

Physical Properties

Description	Specifications	Result
Appearance	Light to moderate golden viscous liquid	Light to moderate golden viscous liquid
Gardner Color – *Orbeco-Hellige Comparator*	3 – 9	N/A
Specific Gravity – *Optima OPD-E*	1.10 – 1.30	N/A
pH (d25°) – *Fisher Accumet AB150*	1.50 – 3.00	N/A
Flash Point (°F) - *Rapid Flash Closed-Cup Tester*	270 - 300	N/A
Infrared IR – *Spectrum Two Perkin/Elmer*	Reference Spectra	PASS

Batch Date: 5-19-2020 Batch # 52020

Shelf Life 5 – 7 years

Shipping Date: 6/25/2020
Shipped to: Microbac Laboratories

Name Arthur V. Martin Ph.D. President

Authorized Signature / Date 6/24/2020

Do NOT store in steel or metal containers. Store in plastic or glass containers only.